541 **Determining HIV-1 Coreceptor Tropism Using PBMC Proviral DNA Derived from Aviremic Blood Samples**

BACKGROUND

- HIV-1 utilizes CD4 and either the CCR5 and/or CXCR4 coreceptor to enter the host cell.
- Agents that block the ability of HIV to utilize CCR5 are available for use in the clinic or in late stage development.
- Determining the tropism of a patient's virus prior to CCR5 antagonist therapy is recommended.
- Current tropism assays are routinely performed using envelope (env) sequences from plasma viral RNA and thus cannot be used to evaluate patients with undetectable levels of plasma virus.
- To address this limitation, we have begun to adapt a pseudovirion-based tropism assay to utilize PBMC-derived env sequences.

METHODS

- Plasma and PBMC samples were obtained from 22 patients in the UCSF SCOPE cohort, CD4 counts, HIV VL and nadir CD4 values are shown for each patient.
- Coreceptor tropism of plasma samples was determined using the Trofile assay.
- Relative ability to infect U87 cells expressing CD4 and CCR5 or CXCR4 is shown for viral pools and clones. The magnitude of infectivity is based on luciferase production in the U87 target cells
- Genomic DNA was extracted from PBMC samples and amplified gp160 *env* sequences were processed through the Trofile assay.
- Multiple *env* clones were isolated from PBMC-associated DNA from 5/12 of the aviremic samples. Tropism and gp160 sequences of env clones were determined.
- Additionally, multiple *env* clones were isolated from four samples with paired plasma and PBMC virus populations and the tropism and gp160 sequences of those env clones were determined.
- gp160 sequences were aligned and NJ phylogenetic trees were constructed with HXB2 sequences as the root.
- Phenotypically determined tropism results from *env* clones were compared to predictions of tropism based on clone V3 sequences (SVM, PSSM and 11/25 rule).

Table 1: Tropism determination utilizing env amplified from PBMC-derived DNA in twelve aviremic HIV+ patients

| Sample ID | Tronism | Magnitude of Infectivity | | CD4 | VI | Nadir | | | | | Average N | lagnitude | V | 3 Tropis | sm |
|-----------|---------|--------------------------|-----|-------|------------|-------|-----------|--------|--|---------|-----------|-----------|------------|------------|------------|
| | порізії | R5 | X4 | - 004 | VL | CD4 | | No. of | | | of Infe | ctivity | F | Predictio | on |
| PBMC 1 | R5 | +++ | - | 711 | 43 | 10 | Sample ID | clones | V3 sequences | Tropism | R5 | X4 | SVM | PSSM | 11/25 |
| _ | | | | | | | | | | | | | | | |
| PBMC_2 | R5 | ++ | - | 930 | <40 | 118 | PBMC_1 | 9 | CTRPNNNTRKGVHIGPGSVWYTTGEIIG-DIRQAHC | R5 | +++ | - | R5 | R5 | R5 |
| | DE | | | 420 | -40 | 269 | | | | | | | | | |
| PBIMC_3 | RD | +++ | - | 420 | <40 | 208 | | 16 | | P5 | | _ | DS | D5 | D5 |
| PBMC_4 | R5 | ++ | - | 383 | <40 | 98 | | 10 | | КJ | TT | _ | КJ | Ŋ | КJ |
| | | | | | | | | | | | | | | | |
| PBMC_5 | R5 | +++ | - | 606 | <40 | 187 | PBMC_8 | 5 | CTRPGNNTRRSITIGPGRAFYTN-NIIG-DIRQAYC | Dual | ++ | + | X4 | R5 | R5 |
| PBMC 6 | R5 | | _ | 274 | ~40 | 27 | | 3 | · · · · · · · · · · · · · · · · · · · | Dual | ++ | + | X4 | R5 | R5 |
| | | | | 214 | NTU | 21 | | 2 | S.KKRVYTG.KS.KF. | X4 | - | ++ | X4 | X4 | R5 |
| PBMC_7 | DM | +++ | ++ | 289 | <40 | 133 | | | | | | | | | |
| | | | | 400 | | | PBMC 9 | 8 | CTRPHNTIRRRIHIGPGRAFYT-GQTTGDIRKAHC | Dual | ++ | +++ | X4 | X4 | X4 |
| PBMC_8 | DM | ++ | ++ | 126 | <40 | 37 | _ | 4 | N NT G Т Т | Dual | | | R5 | R5 | R5 |
| PBMC_9 | DM | ++ | +++ | 264 | <40 | 50 | | 4 | | Dual | | | | V A | |
| | | | | | | | | | ••••••V••••••••••••••••••••••••••••••• | Duai | . | ++ | A 4 | A 4 | A 4 |
| PBMC_10 | DM | +++ | ++ | 290 | <40 | 24 | | 1 | •••••• | R5 | + | - | X4 | X4 | X4 |
| DDMC 11 | DM | | | 202 | -40 | 10 | | | | | | | | | |
| | | TTT | TTT | 203 | <40 | 19 | PBMC_12 | 14 | CTRPSNKTRKKVTLGPGKVWYTTEVKGDIRKAHC | X4 | - | ++ | X4 | X4 | X4 |
| PBMC_12 | X4 | - | +++ | 276 | <40 | 38 | | 1 | v | X4 | - | ++ | X4 | X4 | X4 |

I Table 3: Concordant tropism results in *env* sequences amplified from plasma RNA and **PBMC-associated DNA in ten viremic patients**

| Sample ID | S |
|-----------|---|
| Paired_1 | P |
| Paired_2 | P |
| Paired_3 | P |
| Paired_4 | P |
| Paired_5 | P |
| Paired_6 | P |
| Paired_7 | P |
| Paired_8 | P |
| Paired_9 | P |
| Paired_10 | P |

Table 2: V3 sequence and tropism diversity among *env* clones amplified from PBMC-derived DNA

I Table 4: V3 sequence and tropism diversity among *env* clones amplified from plasma RNA and PBMC-derived DNA in four viremic patients

| urce | Tropism | Magnitude of | Infectivity X4 | CD4 | VL | Nadir CD4 | | No. of | | | Ave Ma of Infe | gnitude ctivity | V3 Tro | opism Pre | diction | No. CI | ones |
|------|-----------|--------------|-------------------|------|---------------------|--------------|-----------|--------|--|-----------|-------------------|--------------------|-----------|-----------|-----------|--------|------|
| | | 110 | Λ 1 | | | | Sample ID | clones | V3 sequences | Tropism | R5 | X4 | SVM | PSSM | 11/25 | Plasma | PBMC |
| ema | R5 | | _ | 1048 | 15 100 | 877 | Paired_3 | 9 | CARPNNNTRKSIHIGPGKAFYTTGSIIGDIRQAHC | R5 | +++ | - | R5 | R5 | R5 | 6 | 3 |
| | | | | 1040 | 10,100 | 011 | | 1 | F | R5 | ++ | - | R5 | R5 | R5 | 1 | 0 |
| SINC | K5 | ++ | - | | | | | 1 | .TG | R5 | +++ | - | R5 | R5 | R5 | 1 | 0 |
| | DE | | | 004 | 40.054 | 500 | | 1 | .TARSAA | R5 | +++ | - | R5 | R5 | R5 | 1 | 0 |
| asma | KO | +++ | - | 821 | 19,251 | 289 | | 8 | .TN | R5 | +++ | - | R5 | R5 | R5 | 1 | 7 |
| BMC | R5 | ++ | - | | | | | 1 | F | R5 | +++ | - | R5 | R5 | R5 | 0 | 1 |
| | | | | | | | | 1 | .v | R5 | +++ | - | R5 | R5 | R5 | 0 | 1 |
| asma | R5 | ++ | - | 453 | 313,000 | 343 | Paired_5 | 11 | CTRPNNNTKKGITMGPGRVYYTTGQIVGDIRRAHC | Dual | +++ | +++ | R5 | X4 | R5 | 11 | 0 |
| BMC | R5 | ++ | - | | | | | 2 | Ү. | Dual | +++ | +++ | X4 | X4 | R5 | 2 | 0 |
| | | | | | | | | 4 | к.І | Dual | ++ | +++ | X4 | X4 | X4 | 0 | 4 |
| asma | R5 | ++ | - | 449 | 90,814 | 449 | | 1 | DY. | Dual | ++ | ++ | X4 | X4 | X4 | 0 | 1 |
| ВМС | R5 | ++ | - | | | | | 1 | K.IY. | X4 | - | ++ | X4 | X4 | X4 | 0 | 1 |
| | | | | | | | Paired 7 | 15 | CTRPNNNTRKGIHIGPGSAFYATGEVTGDIRQAYC | R5 | +++ | - | R5 | R5 | R5 | 9 | 6 |
| asma | DM | ++ | ++ | 40 | 53,600 | 26 | | 2 | GSAIIKF. | R5 | +++ | - | R5 | R5 | R5 | 2 | 0 |
| вмс | DM | ++ | +++ | | | | | 1 | N | R5 | +++ | - | R5 | R5 | R5 | 1 | 0 |
| | | | | | | | | 2 | G | R5 | +++ | - | R5 | R5 | R5 | 0 | 2 |
| asma | R5 | ++ | - | nd | 78,000 | nd | | 1 | A | R5 | +++ | - | R5 | R5 | R5 | 0 | 1 |
| RMC | R5 | | _ | | , | | | 1 | GH. | R5 | +++ | - | R5 | R5 | R5 | 0 | 1 |
| | NJ | *** | | | | | | 1 | AII | R5 | + | - | R5 | R5 | R5 | 0 | 1 |
| sma | R5 | | _ | 431 | 23 700 | 272 | Paired 8 | 7 | CTRRSNNTRKSTHIGEGRALVATGRIIGDIROAHC | R5/Dual | | | R5 | R5 | X4 | 6 | 1 |
| | | | | | 20,100 | <i>L1 L</i> | i unou_o | 4 | | R5 | ++ | - | R5 | R5 | X4 | 3 | 1 |
| | KO | +++ | - | | | | | 2 | A | Dual | +++ | + | R5 | R5 | X4 | 2 | 0 |
| | DE | | | 400 | 00 0 7 5 | 202 | | 1 | K | R5 | ++ | - | R5 | R5 | X4 | 1 | 0 |
| asma | KO | ++ | - | 429 | 22,275 | 393 | | 3 | K | R5 | ++ | - | R5 | R5 | X4 | 1 | 2 |
| BMC | R5 | ++ | - | | | | | 2 | N | R5/Dual | +++ | + | R5 | R5 | X4 | 0 | 2 |
| | | | | | | _ | | 2 | v | R5/Dual | ++ | + | R5 | R5 | X4 | 0 | 2 |
| asma | DM | + | ++ | 11 | 840 | 3 | | 1 | N | Dual | +++ | + | R5 | R5 | R5 | 0 | 1 |
| BMC | DM | +++ | ++ | | | | | 1 | N | Dual | ++ | + | R5 | R5 | X4 | 0 | 1 |
| | | | | | | | | 1 | VK | Dual | ++ | + | R5 | R5 | X4 | 0 | 1 |
| asma | R5 | +++ | - | 453 | 505,000 | 343 | | 1 | N | R5 | ++ | - | R5 | R5 | R5 | 0 | 1 |
| ВМС | R5 | +++ | - | | | | | 1 | G | R5 | +++ | - | R5 | R5 | X4 | 0 | 1 |

Jonathan Toma¹, Arne Frantzell¹, Rebecca Hoh², Jeffrey Martin², Steven G. Deeks², Christos Petropoulos¹, Wei Huang¹ biosciences

¹Monogram Biosciences, Inc., South San Francisco, CA, USA

²University of California, San Francisco, CA, USA



• Figure 1: Diversity of gp160 sequences and tropism in env clones derived from PBMCassociated DNA



Figure 2: Diversity of gp160 sequences and tropism in env clones derived from paired plasma RNA and PBMC-associated DNA





17th Conference on Retroviruses and Opportunistic Infections

Jon Toma Monogram Biosciences, Inc 345 Oyster Point Blvd. South San Francisco, CA 94080 itoma@monogrambio.com

San Francisco, CA **February 16-19th, 2010**

RESULTS

- Tropism was successfully determined in all 22 PBMC samples and in 10/10 viremic plasma samples. The remaining 12 plasma samples had low or undetectable viral loads.
- Of the 12 aviremic patients, six PBMC proviral DNA-derived viruses were R5, five were DM and one was X4 (Table 1).
- Clonal analysis of PBMC-derived *env* in 5/12 aviremic samples shows two samples made up of relatively homogenous R5 clones, one sample entirely composed of closely related X4 clones and two samples with majority dual-tropic clones (Table 2, Figure 1).
- In the 10 viremic patients, PBMC and plasma tropism determinations were concordant (8 R5, 2 DM) (Table 3).
- *env* sequences derived from plasma and PBMC from the same patient exhibited extensive homology. Clonal analysis did not reveal phylogenetic distinctions between plasma and PBMC samples from 3/4 paired samples (Table 4, Figure 2).
- Tropism of clones derived from plasma and PBMC were concordant (Figure 2): one DM virus population was comprised of dual-tropic variants with one X4 clone; one R5 virus population was comprised predominantly of R5 variants with a minor subpopulation of dual-tropic variants which weakly utilized CXCR4 for infection; the two R5 virus populations were comprised of R5 variants.

CONCLUSIONS

- Full-length *env* can be efficiently amplified from cell-associated HIV-1 DNA of patient PBMC to determine coreceptor usage.
- Viral tropism and phylogenetic relatedness of virus populations and clones were generally concordant between paired plasma and PBMC samples.
- The ability to test tropism using cell-associated HIV may be applicable to determining the suitability of CCR5 antagonist use in patients with low or undetectable plasma viral load.

ACKNOWLEDGEMENTS

- We are grateful to the Monogram Biosciences Clinical Reference Lab for their assistance in performing Trofile assays used in this study.
- This study was supported in part by grant funding from NIH UL1 RR024131 and P30 AI027763.