

REVIEWS IN BASIC AND CLINICAL GASTROENTEROLOGY

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Resistance to Direct Antiviral Agents in Patients With Hepatitis C Virus Infection

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Chronic hepatitis C virus (HCV) infection is one of the major causes of cirrhosis, hepatocellular carcinoma, and liver failure that leads to transplantation. The current standard treatment, a combination of pegylated interferon alfa and ribavirin, eradicates the virus in only about 50% of patients. Directly acting antiviral (DAA) agents, which inhibit HCV replication, are in phase 1, 2, and 3 trials; these include reagents that target the nonstructural (NS)3 protease, the NS5A protein, the RNA-dependent RNA-polymerase NS5B, as well as compounds that directly inhibit HCV replication through interaction with host cell proteins. Because of the high genetic heterogeneity of HCV and its rapid replication, monotherapy with DAA agents poses a high risk for selection of resistant variants. We review the parameters that determine resistance, genotypic and phenotypic resistance profiles of DAA agents, and strategies to avoid the selection of resistant variants.

Drug resistance results from variations in drug targets (bacteria, fungi, viruses, tumor cells, and so forth) that affect signal transduction pathways, receptors, drug binding sites, or even host proteins to make the therapeutic agent ineffective. Chronic hepatitis C virus (HCV) infection, which affects more than 180 million patients worldwide and is one of the leading causes of cirrhosis and liver failure, is treated with a combination of pegylated interferon alfa and ribavirin. HCV strains with specific mutations that confer resistance to these drugs have not yet been identified in patients, yet only 50%–60% of patients treated with pegylated interferon alfa and ribavirin achieve a sustained virologic response.¹ This low rate of response can be accounted for by several viral and host-related factors.^{2–5} Furthermore, because of significant side effects associated with the administration of interferon alfa and ribavirin, combination drug doses and treatment duration are restricted. More effective and better-tolerated treatment options for HCV infection are needed. Directly acting antiviral (DAA)

agents that are also named specific, targeted antiviral therapies (STAT-C) for HCV infection are in phase 1–3 trials. We review resistance to DAA agents, focusing on compounds in development such as reagents that target the HCV nonstructural (NS)3 protease, the NS5A protein, and the RNA-dependent RNA polymerase NS5B. We also discuss indirect inhibitors or compounds that inhibit HCV replication by not yet completely resolved mechanisms, such as cyclophilin inhibitors, nitazoxanide, and silibinin (Table 1, Figure 1).

Parameters That Affect Resistance

Heterogeneity of HCV

HCV has a high rate of turnover; its half-life was estimated to be only 2–5 hours, with the production and clearance of 10^{10} to 10^{12} virions per day in an infected patient.^{6,7} The HCV-RNA-dependent RNA-polymerase NS5B has poor fidelity and lacks proof-reading activity. Because of this feature and the high replication activity of HCV, a large number of viral variants are produced continuously during infection (error rate, approximately 10^{-3} to 10^{-5} mutations per nucleotide per genomic replication).⁸ The majority of variants produced result in loss of viral replication via loss of function of encoded proteins or lack of tolerance by the host's immune system.^{9,10} However, a large number of variants survive; the sum of these different isolates in an infected patient is called *viral quasispecies*. Typically a dominant strain (wild-type) is detectable within the viral quasispecies along with strains that are present at lower frequencies. The frequencies of HCV isolates depend on their replication efficacies and other known and unknown viral and host factors. During the phylogenetic evolution of HCV, at least 6 different genotypes and more than 100 subtypes have devel-

Abbreviations used in this paper: DAA, directly acting antiviral; HCV, hepatitis C virus; NS, nonstructural.

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Table 1. Direct Antiviral Drugs in Development for HCV Infection

Drug name	Company	Target/active drug	Study phase
NS3/4A protease inhibitors			
Ciluprevir (BILN 2061)	Boehringer Ingelheim	Active site/macrocyclic	Stopped
Telaprevir (VX-950)	Vertex	Active site/linear	Phase 3
Boceprevir (SCH503034)	Schering–Plough/Merck	Active site/linear	Phase 3
TMC435350	Tibotec/Medavir	Active site/macrocyclic	Phase 2
R7227/ITMN-191	InterMune/Roche	Active site/macrocyclic	Phase 2
MK-7009	Merck	Active site/macrocyclic	Phase 2
BI201335	Boehringer Ingelheim	Active site/macrocyclic?	Phase 2
Narlaprevir (SCH900518)	Schering–Plough/Merck	Active site/linear	On hold
BMS-650032	Bristol–Myers Squibb	Active site	Phase 1
PHX1766	Phenomix	Active site	Phase 1
Nucleoside analogue NS5B polymerase inhibitors			
Valopicitabine (NM283)	Idenix/Novartis	Active site/NM107	Stopped
R7128	Roche/Pharmasset	Active site/PSI-6130	Phase 2
R1626	Roche	Active site/R1479	Stopped
PSI-7851	Pharmasset	Active site	Phase 1
IDX184	Idenix	Active site	Phase 1
Nonnucleoside NS5B polymerase inhibitors (NNI)			
BILB 1941	Boehringer Ingelheim	NNI site 1/thumb 1	Stopped
BI207127	Boehringer Ingelheim	NNI site 1/thumb 1	Phase 2
MK-3281	Merck	NNI site 1/thumb 1	Phase 1
Filibuvir (PF-00868554)	Pfizer	NNI site 2/thumb 2	Phase 2
VCH759	ViroChem Pharma	NNI site 2/thumb 2	Phase 1
VCH916	ViroChem Pharma	NNI site 2/thumb 2	Phase 1
VCH222	ViroChem Pharma	NNI site 2/thumb 2	Phase 1
ANA598	Anadys	NNI site 3/palm 1	Phase 1
HCV-796	ViroPharma/Wyeth	NNI site 4/palm 2	Stopped
GS-9190	Gilead	NNI site 4/palm 2	Phase 1
ABT-333	Abbott	NNI site 4/palm 2	Phase 1
NS5A inhibitor			
BMS-790052	Bristol-Myers Squibb	NS5A domain 1 inhibitor	Phase 1
Indirect inhibitors/unknown mechanism of action			
Debio 025	Debiopharm	Cyclophilin inhibitor	Phase 1
NIM811	Novartis	Cyclophilin inhibitor	Phase 1
SCY-635	Scynexis	Cyclophilin inhibitor	Phase 1
Nitazoxanide		PKR induction?	Phase 2
Silibinin	Rotapharm–Madaus	Polymerase inhibitor?	Phase 2

PKR, double-strand RNA activated protein kinase.

oped that are characterized by large differences in nucleotide sequence (31%–33% and 20%–25%, respectively).¹¹ Thus, the development of HCV therapies is challenged by the different genotypes and subtypes of virus in different patients and also the variation in viral isolates in each patient. Given the heterogeneities in viral sequence, not all drugs will be equally effective for all HCV genotypes, subtypes, and isolates. However, variations are not distributed equally throughout the HCV genome; compounds that target more conserved regions (ie, the active site of the RNA-dependent RNA polymerase) have a better chance of efficacy in patients infected with different genotypes and subtypes.

HCV genotypes and resistance. Many DAA compounds that are being tested in clinical studies are designed to inhibit HCV genotype 1 isolates and the majority of studies have been performed in patients infected with HCV genotype 1. However, telaprevir, an inhibitor of the HCV NS3 protease that targets the active site of

the enzyme, has been given subsequently also to patients infected with genotypes 2, 3, and 4. In these patients, viral load decreased by 3.9, 0.5, and 0.9 log₁₀ IU/mL, respectively, after 2 weeks of monotherapy, compared with 4.4 log₁₀ IU/mL in genotype 1-infected patients.^{12–14} Also, boceprevir has been reported to have weaker antiviral activity in patients infected with genotypes 2 or 3 and similar results are to be expected for other HCV protease inhibitors. Different levels of activity against different HCV genotypes also are expected from nonnucleoside NS5B inhibitors based on results from *in vitro* studies; little clinical data are available.¹⁵ Administration of VCH-759, for example, which binds the thumb 2 site to inhibit the polymerase NS5B, for 10 days led to a mean maximal decrease in HCV RNA log₁₀ IU/mL of 2.5 in genotype 1-infected patients, whereas no viral decline was observed in a genotype 6-infected patient.¹⁶

The active site of NS5B is highly conserved between different genotypes, so nucleoside inhibitors could have

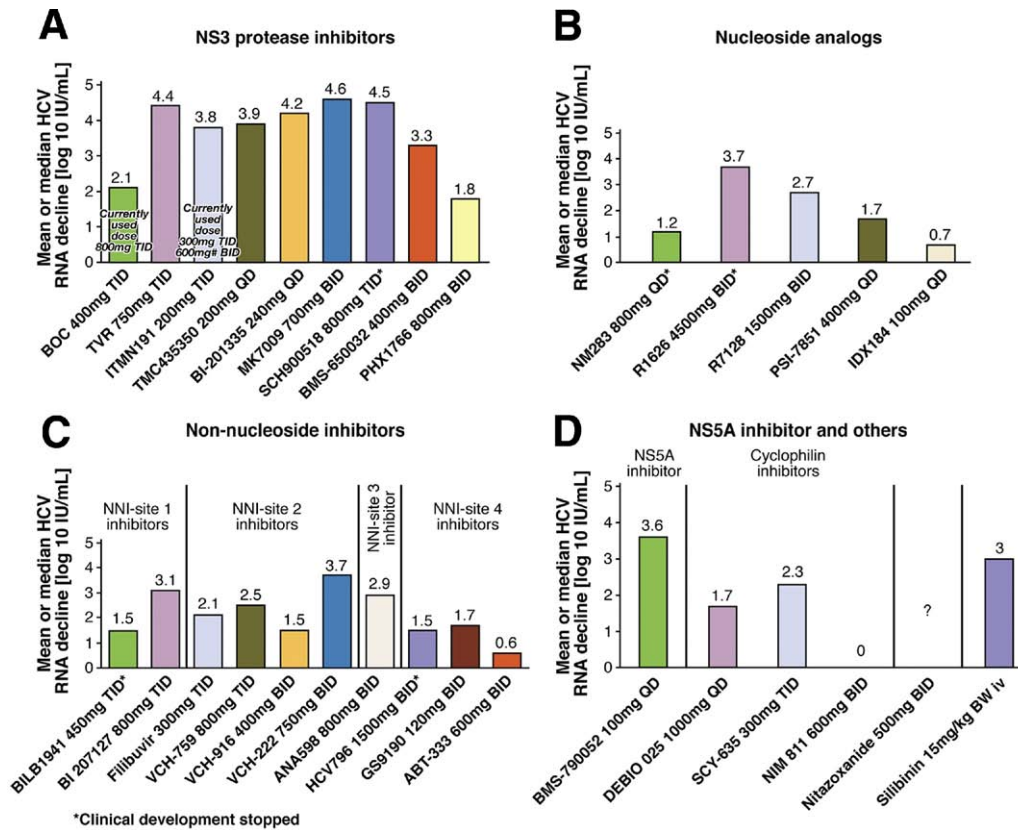


Figure 1. Antiviral activities (mean or median maximum decrease in HCV RNA log₁₀ IU/mL) of DAA agents, as determined from phase 1 monotherapy studies (3–14 days) in patients infected with HCV genotype 1. Different reagents or durations of treatment were not compared directly in these studies. In patients who received the DAA for 7–14 days, the decrease in viral load might not reflect only the activity of the DAA agent but also the emergence of resistant variants. *Clinical development stopped. (A) NS3 protease inhibitors. (B) Nucleoside inhibitors of NS5B. (C) Nonnucleoside inhibitors of NS5B. (D) NS5A inhibitor, indirect inhibitors and agents with unknown mechanism of action. QD, every day, BID, twice a day, TID, three times a day, BW, body weight, #, 900 mg BID stopped from further development due to increase of liver enzymes in single cases.

similar levels of activity in patients infected with different genotypes. However, there are no data on the efficacy of monotherapy with nucleoside analogs against different genotypes. An interim analysis of data from a placebo-controlled trial of patients infected with HCV genotypes 2 or 3 who did not respond to interferon therapy reported increased rapid virologic response rates after 4 weeks of triple therapy with pegylated interferon alfa, ribavirin, and the nucleoside analogue R7128, compared with standard combination therapy (without the NS5B inhibitor). So, it appears that NS5B inhibitors have antiviral activity in patients with HCV genotypes 2 or 3.¹⁷

No clinical data are available on the efficacy of NS5A inhibitors in patients with HCV non-genotype 1, but given the relatively high level of heterogeneity in the NS5A protein, differences in antiviral activities against different genotypes are expected.

Agents that act on host proteins, such as cyclophilin inhibitors, would be predicted to have similar levels of antiviral activity against all HCV genotypes. However, monotherapy with the cyclophilin inhibitor Debio-025 was significantly less effective in patients infected with genotypes 1 or 4 compared with genotypes 2 or 3; viral

loads decreased by 2.5 and 4.4 log₁₀ IU/mL, respectively, after 4 weeks monotherapy with 1000 mg Debio-025.¹⁸ For monotherapy with nitazoxanide, another indirect inhibitor of HCV replication, data on viral load reduction are available only in patients infected with HCV genotype 4.¹⁹ However, indirect evidence for differences between HCV genotypes is available from combination therapies. Triple therapy with nitazoxanide, pegylated interferon, and ribavirin, after a lead-in phase with only nitazoxanide, resulted in different virologic response rates in patients infected with HCV genotype 1 compared with genotype 4 (5%–12% vs 64%, respectively).^{20,21} Finally, silibinin, for which the mechanism of action is not yet entirely resolved, administered intravenously (20 mg/kg/day) for 7 days, led to a mean decline of the HCV-RNA concentration by 3 log₁₀ IU/mL. So far, mainly genotype 1 and single patients with genotype 2, 3, and 4 infections have been investigated and no data on differences of antiviral activities for the different HCV genotypes are available.²²

HCV subtypes and resistance. Differences in the HCV genome sequence are smaller between subtypes than between genotypes, resulting in smaller variations in

antiviral activity of DAA agents between subtypes than genotypes.

However, differences between subtypes have been described in the activities of the linear protease inhibitors telaprevir and boceprevir. In clinical studies of telaprevir alone or in combination with pegylated interferon (with or without ribavirin), selection of resistant variants and viral breakthrough have been observed consistently more frequently in patients infected with HCV subtype 1a than subtype 1b.^{23–25} In the protease inhibition for viral evaluation (PROVE) 1 study (triple therapy: telaprevir, pegylated interferon alfa 2a, and ribavirin), 11 of the 12 patients who experienced a viral breakthrough were infected with HCV subtype 1a; only 1 of the 12 was infected with subtype 1b.²⁵ This difference was shown to result from nucleotide differences at position 155 in HCV subtype 1a (aga, encodes R) vs 1b (cga, also encodes R). The mutation most frequently associated with resistance to telaprevir is R155K; changing R to K at position 155 requires 1 nucleotide change in HCV subtype 1a and 2 nucleotide changes in subtype 1b isolates (Figure 2). Selection of the resistant variant R155K to monotherapy with the NS3 inhibitor boceprevir also differed between patients infected with HCV subtypes 1a and 1b.²⁶ Taken together, based on differences in codon usage for telaprevir and boceprevir several resistant variants were selected typically in subtype 1a- (V36M, R155K/T) or 1b- (R155Q, V170A) infected patients.^{23,26} Differences between subtype 1a- and 1b-infected patients also were observed for macrocyclic NS3 protease inhibitors (ITMN191/R7227, BI201335).^{27,28}

Nucleoside analogs that bind to the highly conserved, active site of NS5B seem to have an equal antiviral activity against different HCV subtypes; no differences have been reported for NM283, R1626, and R7128 in patients infected with HCV subtypes 1a or 1b.^{29–31}

Polymerase inhibitors binding to less-conserved allosteric sites of NS5B have a greater risk of different anti-

viral activities and different resistance profiles in patients infected with different subtypes. Administration of BI207127, a nonnucleoside inhibitor of NS5B that binds to the thumb 1 site (NNI site 1), led to a viral decline in all patients infected with HCV genotype 1, without viral breakthrough during treatment for 5 days. However, BI207127 was reported to be less active against HCV subtype 1a than 1b.³² BILB1941, another thumb 1 site inhibitor, also had greater antiviral activity in patients infected with subtype 1b in comparison with subtype 1a.³³

Filibuvir (PF-00868554) is a thumb 2 site (NNI site 2) inhibitor of NS5B. Differences in resistance to this drug were observed between patients infected with HCV subtypes 1a and 1b after 8 days of monotherapy.³⁴ Also, triple therapy with filibuvir, pegylated interferon alfa 2a, and ribavirin resulted in a rapid virologic response more frequently in patients infected with subtype 1a (14 of 19) vs 1b (3 of 7) virus.³⁵

Furthermore, differences in the reduction in median HCV-RNA levels have been reported between patients with HCV subtypes 1a (1.4–2.5 log₁₀ IU) and 1b (2.6–3.2 log₁₀ IU) who were treated for 3 days with the non-nucleoside inhibitor ANA598, which binds to the palm 1 site (NNI site 3).³⁶ No viral breakthrough was detected during this short-term treatment but the observed differences of the antiviral activity could result from sequence differences between subtypes 1a and 1b in the ANA598 binding site. No host or HCV subtype-specific differences have been reported to affect the antiviral activity of HCV796, a palm 2 site inhibitor (NNI site 4) of NS5B. However, resistance data have been presented only from patients infected with HCV subtype 1a.^{37,38} In vitro different resistant variants in subtype 1a and 1b isolates were selected during incubation with HCV-796.³⁹

HCV isolates and resistance. Even within a given HCV subtype, variations in amino acid sequences are common and can affect the antiviral activity of an antiviral agent and the barrier of resistance.

For example, the pre-existing dominant strain in a patient's quasispecies can sometimes be an HCV variant with reduced sensitivity to NS3 inhibitors that bind to the active site of the protease. In different studies direct sequencing revealed resistant variants at positions V36, T54, R155, D168, and V170 of the NS3 protease in 0.2%–2.2% of patients as the pre-existing dominant strain.^{10,40–43} Patients with the R155K variant had a reduced response to telaprevir compared with patients without this variant and none of the 4 of 570 patients with this variant already present at baseline as the dominant strain achieved a sustained virologic response during treatment in the PROVE 1/2 studies.⁴⁰ Studies using ultra-deep sequencing methods revealed that most amino acid variants that confer resistance to protease inhibitors are detectable within the HCV quasispecies already before treatment, but at low frequencies. Currently, it is un-

NS3 protease amino acid sequence

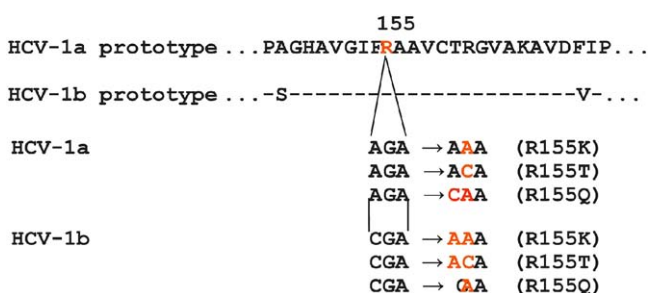


Figure 2. Molecular basis for different probabilities for selection of R155K/T/Q mutations in subtype 1a- vs subtype 1b-infected patients treated with a NS3 protease inhibitor. Selection of R155K/T requires 1 nucleotide exchange in subtype 1a but 2 nucleotide exchanges in subtype 1b patients. Selection of R155Q requires 1 nucleotide exchange in subtype 1b but 2 nucleotide exchanges in subtype 1a patients.

known which frequency of a specific resistant variant influences effectiveness of treatment with an NS3 protease inhibitor.⁴⁴

Mutations that confer in vitro resistance to nucleoside analogs that inhibit NS5B (S96T, S282T, N142T) were not detected in patients with HCV genotype 1 infections.^{10,41,45}

Variants conferring resistance to nonnucleoside inhibitors that bind to different allosteric sites of the NS5B polymerase were detected in 0.6%–2.8% of patients by direct sequencing as pre-existing major variants.^{10,34,41,45,46} However, for nonnucleoside inhibitors resistance mutations also may occur as natural variants more frequently in patients infected with a specific subtype. For example, the C316Y mutation in NS5B is associated most frequently with resistance to HCV-796,³⁷ although C316N also reduces sensitivity to HCV796 in vitro and this variant frequently is observed in many patients infected with HCV subtype 1b.⁴⁵

Resistance mutations selected during treatment with DAA typically are associated with reduced replicative fitness. Interestingly, patients who have not yet received antiviral therapy but are infected with DAA-resistant variants as the pre-existing dominant strain have been reported to have high baseline concentrations of HCV RNA. Thus, it would be interesting to learn how the naturally occurring resistant variants achieved their high replicative capacities in the absence of drug selection.

Barrier to Resistance

The barrier to resistance to a direct antiviral agent is influenced by several parameters such as the HCV genotype and subtype, the activity and pharmacokinetics of the antiviral agent, patient compliance, and the binding affinity of the antiviral compound for its site. We have learned from studies of human immunodeficiency virus and hepatitis B virus (HBV) infection that several recently developed antiviral drugs have a high genetic barrier to resistance because of a flexible binding pattern that allows proper binding even in the presence of resistance mutations⁴⁷ or the requirement of more than 1 amino acid exchange for resistance to occur.⁴⁸ Development of DAA agents for HCV infection still is in its early days with no compounds approved yet and limited studies available on comparative resistance analysis. However, for the agents currently in development, it seems that the genetic barrier of nonnucleoside analogs is lowest, followed by those of NS3 inhibitors, and then nucleoside analogs. For nonnucleoside analogs low to medium antiviral activities (0.6–3.7 log₁₀ HCV RNA IU/mL decline) were observed in clinical studies (Figure 1C). Nucleoside analogs also had relatively low antiviral activities in clinical studies (0.7–1.2 log₁₀ HCV RNA IU/mL decline), which were enhanced up to 3.7 log₁₀ only with very high doses (Figure 1B). NS3 protease inhibitors showed high antiviral activities (up to 4.6 log₁₀ IU/mL HCV RNA

decline) and for the first NS5A inhibitor at low doses a high antiviral activity also was observed (Figure 1D).

Nonnucleoside inhibitors bind to allosteric sites on the surface of NS5B. Here, a high number of variants with multiple amino acid exchanges can occur without functionally altering the NS5B polymerase activity; it is therefore challenging to design an NS5B inhibitor with a high genetic barrier to resistance. Variants that are resistant to most nonnucleoside inhibitors are selected for within a few days of treatment. Moreover, variants with reduced sensitivity to nonnucleoside NS5B inhibitors may be present at high frequencies already before initiation of direct antiviral therapy.

NS3 inhibitors that bind to the active site of the protease are considered to have an intermediate barrier to resistance. Here, variants conferring resistance rarely are observed as naturally occurring variants. However, selection of resistant variants during treatment with an NS3 protease inhibitor followed by a viral breakthrough was observed within 3–4 days of monotherapy.

Nucleoside analogs seem to have the highest genetic barrier to resistance. Mutations at the active site of NS5B most likely cause loss of polymerase function; no resistant variants have been reported to emerge in monotherapy studies (up to 2 weeks) with the nucleoside analogs R1626 or R7128.^{30,31}

In many patients—especially among suboptimal dosing cohorts in phase 1 studies—viral breakthrough occurs, with rapid selection of resistant variants. However, some patients have a continuous decrease in viral load throughout the entire dosing period (2 weeks). Nevertheless, the possibility of long-term suppression of viral replication in patients given monotherapy with a NS3 inhibitor seems unlikely. In one long-term study, telaprevir monotherapy was given to 5 patients for up to 24 weeks. Three patients experienced viral breakthrough with selection of resistant variants, 1 patient discontinued therapy because of side effects, and 1 patient had low levels of HCV RNA in serum at the end of the 24-week treatment period.⁴⁹ No long-term monotherapy studies with NS5B inhibitors have been performed to date and results from additional monotherapy studies with telaprevir from Japan have to be awaited to determine the potential of viral eradication with a single DAA treatment approach.

No clinical data are available on the barrier to resistance for nitazoxanide or silibinin. Interestingly, HCV variants that are resistant to cyclophilin inhibitors have been reported in in vitro studies.^{50,51}

Pharmacokinetics and Compliance

With the exception of silibinin all DAA agents are being developed for oral administration. Both the NS3 inhibitors that are in phase 3 studies, boceprevir and telaprevir, have a relatively short half-life, requiring a 3-times-per-day dosing interval.^{12,52} Trough plasma levels

were correlated with antiviral responses in phase 1 studies of these agents. Moreover, 6 of 10 patients who received 2 daily doses of 1250 mg telaprevir had viral breakthrough and resistance, compared with 1 of 8 patients who received 3 daily doses of 750 mg telaprevir (the subsequent standard therapy) in a phase 1 monotherapy study.²³ Interim analysis after 12 weeks of triple therapy with telaprevir, pegylated interferon, and ribavirin showed no differences in the decrease of antiviral activity or viral breakthrough in patients treated 2 times daily with 1125 mg, compared with 3 times daily with 750 mg telaprevir. However, small numbers (39–42 patients) were enrolled in each cohort and differences might become visible only in larger studies, especially in nonresponders to previous pegylated interferon/ribavirin combination therapy.⁵³

For patients with human immunodeficiency virus infection, compliance is one of the major factors that affect the development of resistance.⁵⁴ Treatment failure from drug resistance has been decreased by reducing the administration of antiretroviral therapy from 3-times-daily to once-daily highly active antiretroviral therapy. Many DAA agents currently in phases 1 and 2 trials have a half-life that allows for once- or twice-daily dosing. Similar to human immunodeficiency virus treatment, ritonavir also is given to HCV patients along with several HCV NS3 inhibitors (ITMN191/R7227, SCH900518) to improve pharmacokinetic properties and prolong dosing intervals. However, about 10% of patients are intolerant to ritonavir because of its side effects. Thus, the addition of ritonavir to triple therapies of DAA, pegylated interferon, and ribavirin may enhance early discontinuation rates and thereby may reduce a patient's potential to achieve a sustained virologic response.

Combination Therapies

In human immunodeficiency virus and HBV infection combination therapies are used to overcome rapid selection of variants that are resistant to single agents and increase the odds of long-term suppression of viral replication. HCV variants with amino acid changes that confer resistance to DAA compounds retain their sensitivity to interferon alfa and non-cross-resistant drug classes *in vitro*.^{55–57}

DAA plus pegylated interferon. Combination therapies of protease inhibitors with pegylated interferon alfa have been tested for their abilities to overcome resistance to single agents and for their additive or even synergistic antiviral activities. In a phase 1 study of 14 days of combination therapy with telaprevir and pegylated interferon alfa 2a, a continuous decline in HCV-RNA concentrations, without viral breakthrough, was observed. Furthermore, antiviral efficacy was at least additive, with a maximum decline of 5.5 log₁₀ for the combination therapy, compared with 4.4 log₁₀ for telaprevir monotherapy.⁵⁸ Another study showed that combination therapy with boceprevir with pegylated inter-

feron alfa 2b principally may be effective in patients who did not respond to previous interferon alfa-based treatment regimens.⁵² Combination therapies with pegylated interferon and other protease or polymerase inhibitors had similar effects. However, in the first phase 2 trial, which included a longer duration of therapy, up to 24% of patients who received combination therapy with telaprevir and pegylated interferon alfa 2a experienced a viral breakthrough with emergence of resistant HCV variants. In addition, a high relapse rate (48%) was observed.^{25,59}

Triple therapy DAA plus pegylated IFN plus ribavirin. The addition of ribavirin to combination therapy with a protease inhibitor and pegylated interferon alfa significantly increased antiviral activity, lowered relapse rates, and reduced viral breakthrough and selection for resistant variants. In a study that compared combination therapy with telaprevir and pegylated interferon alfa to triple therapy that included ribavirin, HCV-RNA levels were undetectable after 12 weeks in 62% and 80% of patients, respectively. Furthermore, viral breakthrough occurred during treatment in 24% of patients on double therapy and 1% on the triple therapy; virologic relapse occurred in 42% and 25% of the patients, respectively.⁵⁹ Similar results were observed in other studies comparing dual vs triple therapies with the NS3 protease inhibitor boceprevir or the polymerase inhibitor R1626.^{30,60–62} However, no studies have been conducted to test combination therapies with DAA and ribavirin but without interferon alfa. Future studies are needed to determine whether ribavirin exerts its antiviral activity only in the presence of interferon alfa or also if it can decrease viral load and prevent selection of resistant variants in combination with a DAA agent.

DAA combination therapies. Another possibility to suppress viral replication, avoid the selection of resistant variants and viral breakthrough, as well as potentially eradicate the virus are DAA combination regimens. Studies in the *in vitro* HCV replicon model and in chimpanzees have indicated the potential of combination therapies with protease and polymerase inhibitors. Indeed, using this approach, HCV was eradicated in a chimpanzee.^{57,63} Results of the first clinical study of the combination of a polymerase inhibitor (R7128) and an NS3 inhibitor (R7227/ITMN191) were presented recently (INFORM-1 study). In this proof-of-principle study, patients were given both DAA agents for up to 2 weeks. HCV-RNA concentrations decreased by as much as 5.2 log₁₀ IU/mL, no viral breakthrough was observed, and HCV RNA was undetectable by the end of the study in up to 63% of patients, measured using a highly sensitive assay.⁶⁴ The potential of this combination therapy to suppress replication for longer durations, without viral breakthrough or selection of resistant variants, and to ultimately achieve a sustained virologic response, has to be shown in future studies.

Characterization of Resistant Variants Selected During Treatment With DAA

The resistance profile of an HCV quasispecies is best determined by analysis of the genome sequence that is targeted by the DAA agent using a highly sensitive clonal sequencing approach. By direct sequencing, only amino acid variations that occur in the HCV quasispecies at a frequency of approximately 25% can be detected, whereas cloning and sequencing of, for example, 80 isolates, viral variants present at approximately 5% of the population will be detected. In addition, the clonal approach also allows analysis of mutations linked on a given isolate. Moreover, because of the rapid viral turnover during HCV infection and because highly resistant variants typically are associated with a significantly impaired replicative fitness these variants will be overgrown by wild-type virus shortly after termination of antiviral therapy and/or compensated by additional mutations that enhance the replication efficiency. The likeliness of selection of isolates with compensated replicative fitness increases with the duration of DAA therapy without complete suppression of viral replication. Furthermore, for combination therapies resistant viral variants may be selected in the first days during treatment followed by a continuous viral decline because of a lack of cross-resistance between the different antiviral drugs.

It is not known whether resistant variants can persist in the long term as major or minor quasispecies in nonresponsive patients or those with viral breakthrough or relapse after DAA therapy. Thus, genotypic sequence analysis should be performed in the first days during therapy, directly after viral breakthrough, at the end of treatment, and during short-term and long-term follow-up evaluation. Initial characterization of the HCV resistance profile to DAA agents, using the highly sensitive clonal sequencing method, is mandatory. However, other methods for sequence analysis are available (ie, direct sequencing, TaqMAMA, hybridization assays, restriction enzyme assays, deep sequencing methods). It remains to be established which approach will be most useful to identify DAA agent resistance mutations in clinical practice.⁶⁵

The first characterization of HCV resistance variants selected during monotherapy with telaprevir and identified by the clonal method identified mutations that were not predicted with the HCV replicon assays.^{23,66} Previous *in vitro* replicon studies identified the A156S/V/T mutation in NS3, but not mutations at V36, T54, or R155, which were selected in patients treated with telaprevir. High doses of telaprevir were used in the *in vitro* replicon studies, whereas lower concentrations of boceprevir selected for the T54A and V170A mutations in NS3 in the HCV replicon system. Mutations at position Q41 and F43 of NS3 were identified in the replicon assay, but these variants were not selected in patients treated with telaprevir or boceprevir so far.⁶⁷ Mutations in NS3 that

conferred resistance to boceprevir, such as V55A, also were not predicted from *in vitro* studies but were detected using highly sensitive genotypic analysis of the HCV quasispecies in patients from the phase 1 monotherapy study.²⁶

Interestingly, for both telaprevir and boceprevir during monotherapy a large number of mutational variants were selected ($n = 25-31$) and restrictions (increase of more than 5% frequency from baseline, detection in more than 1 patient, decline after the end of treatment) were made for further analysis.^{23,26,68} Finally, mutations at 6 amino acid positions (V36, T54, V55, R155, A156, and V170) were associated clearly with resistance to these protease inhibitors. A potential importance of the multiple additional low-frequency variants at other positions has to be investigated in future studies.

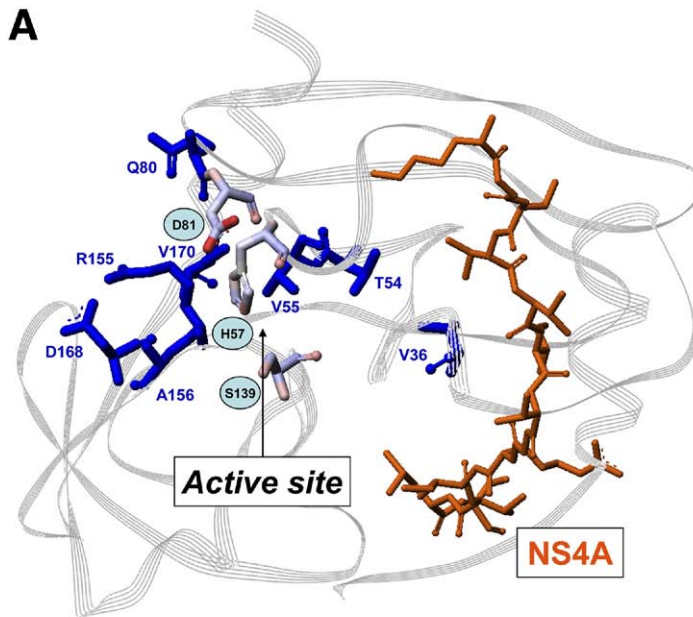
After genotypic characterization, levels of resistance to the antiviral agent used in the clinical study, and potential cross-resistance to other direct antivirals must be determined by phenotypic assays. Different replicon- and enzyme-based phenotypic assays have been used to determine the IC_{50}/EC_{50} values of DAA agents for HCV replication or enzymatic functions of wild-type and mutant HCV proteins.^{23,26,27,69,70} The majority of replicon assays is based on the HCV con1b isolate with restrictions for variants detected in subtype 1a isolates and other genotypes.³⁹ Furthermore, typically only the detected variant, alone or in combination with another resistant variant, rather than the entire NS3 protease gene from a given patient, is introduced into NS3 for analysis in a phenotypic assay.

The data from clonal genotypic and phenotypic resistance analyses of telaprevir and boceprevir from phase 3 trials have been published.^{23,24,26} However, the resistance profiles of the majority of DAA agents have not been characterized fully. We summarize the available resistance genotypic and phenotypic resistance profile of DAA agents that are in phase 1, 2, or 3 trials.

NS3 Protease Inhibitors

BILN2061 (Ciluprevir) was the first NS3 protease inhibitor investigated in clinical studies. BILN2061 showed a high antiviral activity in genotype 1 and a variable antiviral activity in genotype 2/3-infected patients after dosing for 2 days. However, further development of BILN2061 was stopped and no clinical resistance data are available.⁷¹⁻⁷³

Telaprevir. Telaprevir has a high antiviral activity (Figure 1A) but a medium genetic barrier to resistance.¹² During monotherapy of genotype-1-infected patients with different doses of telaprevir for 14 days mainly viral variants at 6 positions within the NS3 protease were selected rapidly (Figure 3).²³ These variants were detected either alone or in combination on the same isolate (mainly positions 36+155 or 36+156). Phenotypic analysis showed low- to medium-level (V36, T54, R155,



B

	V36A/M	T54S/A	V55A	Q80R/K	R155K/T/Q	A156S	A156T/V	D168A/V/T/H	V170A/T
Telaprevir (linear)			*						*
Boceprevir (linear)							*		
SCH900518 (linear)									
BILN-2061 ** (macrocylic)									
ITMN191/R7227 (macrocylic)						*	*		
MK-7009 (macrocylic)									
TMC435350 (macrocylic)									
BI-201335 (macrocylic?)									

* Mutations associated with resistance in vitro but not described in patients so far

** Mutations associated with resistance in vitro

Figure 3. Amino acid positions within the NS3/4A protease associated with (A) resistance mutations to different NS3 protease inhibitors and a (B) cross-resistance table of different NS3 protease inhibitors based on mutations selected in patients from clinical studies and/or from in vitro studies. Amino acids forming the catalytic triad (H57, D81, S139) are highlighted. *Mutations associated with resistance in vitro but were not described in patients. **Mutations associated with resistance in vitro. Resistance mutations of linear NS3 protease inhibitors are shown in blue and resistance mutations described for macrocylic NS3 protease inhibitors are shown in green.

A156S) and high-level (A156V/T) resistant variants. Molecular-based mechanisms of resistance have been proposed.^{66,74–76} In more than 80% of the isolates of the HCV quasiespecies at the end of treatment resistance mutations were detected. High-level resistant variants typically were observed in patients with higher telaprevir exposure and more pronounced viral decline. Generally, an inverse correlation between the level of telaprevir resistance conferred by a single mutation and replicative viral fitness was observed. Double mutants (V36A/M + R155K/T) seem to have a compensatory effect with an increase of replicative fitness together with a relatively high level of resistance.^{23,24} After termination of treatment with telaprevir the number of wild-type isolates rapidly increased

within the first 7–10 days of follow-up evaluation. After 3–7 months no high-level resistance was observed anymore and the majority of resistant variants were replaced by wild-type virus.²³ However, in single patients V36 and R155 variants still were detectable at low levels and in a recent report on long-term follow-up evaluation even after up to 3 years in single patients low- to medium-levels of V36 and R155 variants were observed.⁷⁷

During combination therapy of telaprevir with pegylated interferon alfa A156T/V variants were detectable in the initial viral decline phase but viral levels continued to decline.²⁴ During triple therapy with telaprevir, pegylated interferon alfa 2a, and ribavirin in 3% of the patients (5 of 163) a viral breakthrough with wild-type virus in 1 pa-

tient but high-level resistant variants in 4 patients (V36M+R155K, A156T) was observed. Furthermore, in patients with relapse after the end of treatment also low- and high-level resistant variants with R155K as the main resistance mutation were detected.⁵⁹

Generally, the likeliness of a viral breakthrough is correlated with the response to pegylated interferon alfa/ribavirin combination therapy. The majority of patients with a virologic null-response (<1 log₁₀ decline at week 4 or <2 log₁₀ decline at week 12) to pegylated interferon alfa/ribavirin combination therapy who were retreated with triple therapy including telaprevir showed a viral breakthrough. In partial nonresponders (>2 log₁₀ decline at week 12 but HCV-RNA positive at week 24) only several viral breakthroughs were observed during triple therapy and in patients with a prior relapse after standard combination therapy no viral breakthrough was observed during re-treatment with triple therapy including telaprevir.⁷⁸ Thus, for patients with null-response to pegylated interferon alfa/ribavirin, triple therapy is a form of functional monotherapy with the direct antiviral drug and this is associated with a high probability of viral breakthrough. Future studies have to investigate the possibility to select the optimal patients for triple therapy perhaps with a lead-in phase to characterize the responsiveness to pegylated interferon alfa/ribavirin alone or together with other response predictors (baseline viral load, IL28 polymorphism. This would on the one hand enable the possibility to avoid viral breakthrough with selection of resistant variants and perhaps on the other hand for patients with favorable response predictors no need of the addition of a DAA or very short triple therapies.

Telaprevir was investigated also in genotype 2-, 3-, and 4-infected patients with lower (genotype 2) or virtually absent (genotype 3, 4) antiviral activities.^{13,14} Data on resistant variants selected in patients infected with genotype 2, 3, and 4 isolates will be presented at an upcoming meeting.

Boceprevir. Boceprevir has a medium antiviral activity (Figure 1A) and a medium genetic barrier to resistance.⁷⁹ During monotherapy with 400 mg boceprevir 2 or 3 times daily for 14 days, mainly viral variants at 6 positions within the NS3 protease conferring low- to medium-level resistance were observed (Figure 3). As for telaprevir and also for boceprevir an increasing number of selected variants correlated with a higher viral decline. Interestingly, the mean frequency of wild-type variants was still 86% at the end of treatment and the high-level resistant variant A156T was not observed.²⁶ The currently used dose of boceprevir in the phase 3 trials is 800 mg 3 times daily. However, no monotherapy data with this dose are available and it is unknown whether with the higher dose also high-level resistant variants will be selected.

Genotypic analyses show a largely overlapping cross-resistance profile of boceprevir and telaprevir (Figure 3). However, for example, the V170A variant conferred a higher level of resistance to boceprevir in comparison with telaprevir and this variant was not described to be a frequent variant in patients who received telaprevir.^{23,24,26} Shortly after the end of boceprevir treatment the frequency of wild-type variants increased to 96%. However, in a number of patients 1 year after the end of boceprevir dosing and in single patients even 4 years after the end of boceprevir dosing, either alone or in combination with pegylated interferon, resistant variants still were detected in the HCV quasispecies by clonal sequence analysis.⁸⁰ Boceprevir also was investigated in genotype 2- and 3-infected patients. However, antiviral activity was reduced in these genotypes and no resistance analysis is available from these studies.

Triple therapy with boceprevir, pegylated interferon alfa 2b, and ribavirin in a large phase 2 study in genotype 1 treatment-naïve patients for 2848 weeks was associated with viral breakthrough and selection of resistant variants in 7%–12% of the patients.⁶¹ Patients who received a lead-in treatment with pegylated interferon and ribavirin alone for 4 weeks before the addition of boceprevir for another 24 or 44 weeks showed lower viral breakthrough rates (4%–5%).⁶¹ However, to answer the question whether a lead-in phase with peginterferon and ribavirin alone will generally reduce the probability of a viral breakthrough, analysis of the phase 2 and 3 studies with boceprevir, telaprevir, and other DAA have to be awaited.

Other NS3 protease inhibitors (R7227ITMN191, MK7009, BI201335, TMC435350, SCH900518, BMS-650032, PHX1766). Several NS3 protease inhibitors currently in phase 1/2 development showed high antiviral activities in monotherapy studies (Figure 1A)^{81–87} and initial resistance analyses have been presented at scientific meetings.^{27,28,84,85,88,89} Taken together, differences between linear tetrapeptide and macrocyclic inhibitors binding to the active site of the NS3 protease have been revealed (Figure 3). However, R155 is the main overlapping position for resistance, and different mutations at this amino acid site within the NS3 protease confer resistance to all protease inhibitors that are currently in advanced clinical development (Figure 3).

Polymerase Inhibitors

Several nucleoside inhibitors targeting the active site of the HCV-RNA-dependent RNA-polymerase as well as a number of nonnucleoside inhibitors binding to different allosteric sites of the NS5B polymerase are currently in clinical development (Table 1).

Nucleoside analogs. Valopicitabine (NM283, 2'-C-methylcytidine/NM107), the first nucleoside inhibitor investigated in patients with chronic hepatitis C, showed a low antiviral activity (Figure 1B).²⁹ Because of gastrointestinal side effects the clinical development of NM283

Figure 4. Amino acid positions within the NS5B polymerase associated with (A) resistance mutations to different nucleoside and nonnucleoside inhibitors and a (B) cross-resistance table of different NS5B polymerase inhibitors based on mutations selected in patients or in vitro. Amino acids forming the catalytic triad (D318, D319, D220) are highlighted. *Mutations associated with resistance in patients. **Mutations associated with resistance in vitro. Mutations of 2'-C-methylcytidine and 4'-azidocytidine nucleoside inhibitors are shown in red and pink, respectively. For NNI-site 1, 2, 3, and 4 inhibitors resistance mutations are shown in orange, dark blue, dark green, and yellow, respectively.

was stopped. Very sparse clinical data on resistance are available but from in vitro assays S282T mutation in the NS5B polymerase is known to confer resistance to NM283 (Figure 4).⁹⁰

The second nucleoside inhibitor to be reported in patients with chronic hepatitis C was R1626 (4'-azidocytidine/PSI-6130). A phase 1 study in genotype 1-infected patients observed a high antiviral activity at high doses of

R1626 in genotype 1–infected patients (Figure 1B).³⁰ No viral breakthrough with selection of resistant variants was reported from monotherapy or combination studies with pegylated interferon ± ribavirin.^{30,62} In HCV replicon studies S96T mutation within the NS5B polymerase conferred resistance to R1626.⁹⁰ Because of severe lymphopenia and infectious disease adverse events further development of R1626 was stopped.

R7128, another nucleoside polymerase inhibitor with antiviral activity in genotype 1–infected patients in phase 1 monotherapy studies,⁹¹ currently is being investigated in phase 2 clinical trials in genotype 1–, 2–, and 3–infected patients in combination with pegylated interferon and ribavirin as well as in combination with the protease inhibitor R7227/ITMN191. Also from these studies no viral breakthrough with selection of resistant variants was reported. In vitro, different resistant variants for R7128/NM283 (S282T) and R1626 (S96T) have been selected and cross-resistance was excluded (Figure 4).⁹⁰

Recently, 2 other polymerase inhibitors (PSI-7851 and IDX184) were evaluated in phase 1 clinical trials in patients with chronic hepatitis C (Figure 1B). However, no data on resistance have been presented for these drugs so far.^{92,93}

Nonnucleoside analogs. At least 4 different allosteric binding sites have been identified for inhibition of the NS5B polymerase by nonnucleoside inhibitors (Table 1). A number of mutations conferring resistance to the different chemical classes of nonnucleoside inhibitors have been observed mainly in in vitro studies (Figure 4).

NNI-site 1 inhibitors (thumb 1/benzimidazole site). BILB1941, BI207127, and MK-3281 are NNI-site 1 inhibitors that have been investigated in clinical phase 1 trials with low to medium antiviral activities (Figure 1C).^{32,33,94,95} No viral breakthrough with selection of resistant variants has been reported during 5 days of treatment with BILB1941 or BI207127.^{32,46} However, preexisting NS5B substitutions known to reduce sensitivity were identified for BILB1941 (V494I/A, I424V, P496A) and resistance mutations of NNI-site 1 inhibitors were characterized in vitro (Figure 4).^{96,97}

NNI-site 2 inhibitors (thumb 2/thiophene site). Flibuvir (PF-00868554) is a NNI-site 2 inhibitor with medium antiviral activity in a phase 1 study (Figure 1C).⁹⁸ Preexisting variants conferring reduced sensitivity to flibuvir were observed in single patients (M426A). Several flibuvir resistant variants have been described in HCV replicon studies.⁹⁹ During treatment a high number of different variants was observed. However, variants at position M423 known to confer resistance to flibuvir from in vitro studies were selected in the majority of patients (Figure 4).^{34,99} In a subsequent triple-therapy trial with flibuvir, pegylated interferon alfa 2a, and ribavirin for 4 weeks, a viral breakthrough was observed in 5 of 26 patients.³⁵

VCH-759, VCH-916, and VCH-222 are 3 other NNI-site 2 inhibitors with antiviral activities in monotherapy studies (Figure 1C).^{16,100,101} For VCH-759 as well as VCH-916 viral breakthroughs with selection of resistant variants conferring high (M423T/V/I) and medium (L419V/M, I482L/V/T, V494A/I) levels of resistance were observed (Figure 4).^{16,102} In the majority of patients who experienced a viral breakthrough resistance mutations were found to remain present up to 13 days despite a high HCV replication rate and a reduced replication fitness of the variants in vitro.¹⁶

NNI-site 3 inhibitors (palm 1/benzothiadiazine site). ANA598 is a NNI-site 3 inhibitor that displayed antiviral activity during treatment of genotype 1–infected patients (Figure 1C).³⁶ No viral breakthrough was observed during this short-term monotherapy trial. However, several variants were described from in vitro replicon studies to confer resistance to ANA598 (Figure 4).¹⁰³

NNI-site 4 inhibitors (palm 2/benzofuran site). Monotherapy with the NNI-site 4 inhibitor HCV-796 showed low antiviral activity in genotype 1–infected patients (Figure 1C).³⁸ Viral breakthrough was associated with selection of resistant variants that conferred medium to high levels of resistance in phenotypic analysis^{37,104} (Figure 4). For GS-9190 low antiviral activity was observed in a clinical study and variants conferring resistance were identified in the β -hairpin of the polymerase.^{94,105} ABT-333, another palm site inhibitor, showed antiviral activity in patients with genotype 1 infection (Figure 1C) and from in vitro replicon studies C316Y and Y448C were observed as main resistance mutations.^{106,107} In addition, S556G was reported to be associated with viral breakthrough in a clinical study (Figure 4).¹⁰⁶

NS5A Inhibitor

In a single ascending dose study it was shown that inhibition of the NS5A protein with BMS-790052 leads to a sharp initial decline of HCV-RNA concentrations (Figure 1D).¹⁰⁸ BMS-790052 is the first NS5A inhibitor binding to domain I of the NS5A protein, which was shown to be important for regulation of HCV replication.¹⁰⁹ No clinical data on resistance to this class of drugs have been presented yet and results of multiple dose studies have to be awaited.

Cyclophilin Inhibitors

Cyclophilins are functional regulators of the HCV NS5B polymerase.¹¹⁰ Debio-025, NIM811, and SCY-635 are cyclophilin A inhibitors, which block HCV replication in vitro.^{50,111,112} For Debio-025 antiviral activities in patients infected with different HCV genotypes (1–4) was shown during monotherapy and in combination studies with pegylated interferon (Figure 1D).^{18,113} No viral breakthrough was observed in these studies. However, selection of HCV variants with mutations clustering in the NS5A gene resistant to Debio-025 was possible in the

HCV replicon system.^{51,112} Another cyclophilin inhibitor SCY-635 also showed antiviral activity in genotype 1-infected patients (Figure 1D).⁵⁰ During this 14-day monotherapy study no viral breakthrough was reported. In vitro mutations within the NS5A (T17A, E295K, V444A) and NS5B (T77K, I432V) protein conferred resistance to SCY-635.¹¹⁴ No viral decline in genotype 1-infected patients treated with NIM811 alone was observed in a phase 1 study (Figure 1D).¹¹⁵ However, selection of cell lines conferring resistance to NIM811 without disclosure of the mutations observed was shown in the replicon system.¹¹¹ Taken together, cyclophilin inhibitors may have the potential of broad antiviral activity to different HCV genotypes. However, selection of resistant variants is possible and the natural- and treatment-induced frequency of mutations conferring resistance to the different cyclophilin inhibitors have to be explored.^{116,117}

Nitazoxanide

Nitazoxanide is a thiazolide anti-infective with activity against anaerobic bacteria, protozoa, and viruses. It is approved for treatment of cryptosporidiosis and lambliaosis in the United States. In vitro nitazoxanide was shown to inhibit HCV replication and the mechanism of action was proposed to be induction of double-strand RNA activated protein kinase.¹¹⁸ In a placebo-controlled study monotherapy with nitazoxanide for 24 weeks led to a mean reduction of HCV-RNA viral load in genotype 4-infected patients by 1.55 log₁₀ IU/mL with 30% of patients having undetectable HCV RNA at the end of treatment.¹⁹ Studies in genotype 1-infected patients with a lead-in monotherapy with nitazoxanide for 4 weeks have been performed. However, no virologic response data in genotype 1-infected patients during monotherapy with nitazoxanide have been reported (Figure 1D).^{21,119} Patients experiencing viral breakthroughs were not reported from monotherapy studies. However, in vitro studies suggested the potential of resistance to nitazoxanide most likely was conferred by changes in host proteins and not by mutations in the virus.¹¹⁸

Silibinin

The mechanism of action of silibinin is unknown but inhibition of NS5B polymerase as well as antiviral and anti-inflammatory activities shown in NS5B polymerase assays, in human peripheral blood mononuclear cells, and the HCV replicon, respectively, have been reported.^{120,121} Monotherapy studies with daily intravenous application of silibinin in genotype 1-, 2-, and 3-infected patients for up to 14 days showed good antiviral activities (Figure 1D).²² Studies in larger cohorts of patients including resistance analysis are underway.

Conclusions

The development of DAA agents has increased the number of therapeutic options for patients with HCV

infection. Data from phase 2 studies have shown that rates of sustained virologic response significantly improve in patients given triple therapies of a DAA agent, pegylated interferon, and ribavirin. However, these approaches also have many restrictions and raise some new problems and questions. Because of the high level of genomic heterogeneity among HCV subtypes and isolates, DAA agents will not have similar levels of activities in all patients. Most reagents are being developed to treat patients infected with HCV genotype 1. Although sustained virologic response rates are increasing among these patients, triple therapy is not effective in all patients infected with genotype 1 because of side effects, nonresponsiveness to pegylated interferon or ribavirin, or development of resistance. It is important to increase our understanding of mechanisms of resistance mutations, viral breakthrough, and relapse after therapy with DAA agents. Improving our knowledge of these processes will require more precise and earlier characterization of resistance profiles of patients given monotherapy, long-term follow-up data, and studies with combination therapies. Studies also are underway to test the effects of combinations of DAA agents without interferon alfa or ribavirin—these trials will provide valuable information about the ability of DAA agents to suppress HCV replication without development of resistance or even to eradicate the virus.

References

1. Zeuzem S, Berg T, Moeller B, et al. Expert opinion on the treatment of patients with chronic hepatitis C. *J Viral Hepat* 2009;16:75–90.
2. Wohnsland A, Hofmann WP, Sarrazin C. Viral determinants of resistance to treatment in patients with hepatitis C. *Clin Microbiol Rev* 2007;20:23–38.
3. Ge D, Fellay J, Thompson AJ, et al. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature* 2009;461:399–401.
4. Suppiah V, Moldovan M, Ahlenstiel G, et al. IL28B is associated with response to chronic hepatitis C interferon-alpha and ribavirin therapy. *Nat Genet* 2009;41:1100–1104.
5. Tanaka Y, Nishida N, Sugiyama M, et al. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 2009;41:1105–1109.
6. Neumann AU, Lam NP, Dahari H, et al. Hepatitis C viral dynamics in vivo and the antiviral efficacy of interferon- α therapy. *Science* 1998;282:103–107.
7. Herrmann E, Neumann AU, Schmidt JM, et al. Hepatitis C virus kinetics. *Antivir Ther* 2000;5:85–90.
8. Bartenschlager R, Lohmann V. Replication of hepatitis C virus. *J Gen Virol* 2000;81:1631–1648.
9. Rauch A, James I, Pfafferott K, et al. Divergent adaptation of hepatitis C virus genotypes 1 and 3 to human leukocyte antigen-restricted immune pressure. *Hepatology* 2009;50:1017–1029.
10. Gaudieri S, Rauch A, Pfafferott K, et al. Hepatitis C virus drug resistance and immune-driven adaptations: relevance to new antiviral therapy. *Hepatology* 2009;49:1069–1082.
11. Simmonds P, Bukh J, Combet C, et al. Consensus proposals for a unified system of nomenclature of hepatitis C virus genotypes. *Hepatology* 2005;42:962–973.
12. Reesink HW, Zeuzem S, Weegink CJ. Rapid decline of viral RNA in hepatitis C patients treated with VX-950: a phase Ib, placebo-

- controlled, randomized study. *Gastroenterology* 2006;131:997–1002.
13. Foster GR, Hezode C, Bronowicki JP, et al. Activity of telaprevir alone or in combination with peginterferon alfa-2a and ribavirin in treatment-naïve genotype 2 and 3 hepatitis-C patients: interim results of study C209. *J Hepatol* 2009;50(Suppl 1):S22.
 14. Benhamou Y, Moussalli J, Ratziv V, et al. Results of a proof of concept study (C210) of telaprevir monotherapy and in combination with peginterferon alfa-2a and ribavirin in treatment-naïve genotype 4 HCV patients. *J Hepatol* 2009;50(Suppl 1):S6.
 15. Herlihy KJ, Graham JP, Kumpf R, et al. Development of intergenotypic chimeric replicons to determine the broad-spectrum antiviral activities of hepatitis C virus polymerase inhibitors. *Antimicrob Agents Chemother* 2008;52:3523–3531.
 16. Cooper C, Lawitz EJ, Ghali P, et al. Evaluation of VCH-759 monotherapy in hepatitis C infection. *J Hepatol* 2009;51:39–46.
 17. Gane EJ, Rodriguez-Torres M, Nelson DR, et al. Antiviral activity of the nucleoside polymerase inhibitor R7128 in HCV genotype 2 and 3 prior non-responders: interim results of R7128 1500mg BID with peg-IFN and ribavirin for 28 days. *Hepatology* 2008;48(Suppl):1024A.
 18. Flisiak R, Feinman SV, Jablkowski M, et al. The cyclophilin inhibitor Debio 025 combined with PEG IFNalpha2a significantly reduces viral load in treatment-naïve hepatitis C patients. *Hepatology* 2009;49:1460–1468.
 19. Rossignol JF, Kabil SM, El-Gohary Y, et al. Clinical trial: randomized, double-blind, placebo-controlled study of nitazoxanide monotherapy for the treatment of patients with chronic hepatitis C genotype 4. *Aliment Pharmacol Ther* 2008;28:574–580.
 20. Rossignol JF, Elfert A, El-Gohary Y, et al. Improved virologic response in chronic hepatitis C genotype 4 treated with nitazoxanide, peginterferon, and ribavirin. *Gastroenterology* 2009;136:856–862.
 21. Bacon B, Shiffman M, Lim J, et al. Phase 2 randomized, double-blind, placebo-controlled study of nitazoxanide plus peginterferon and ribavirin in HCV genotype 1 patients: interim analysis shows increase in EVR. *J Hepatol* 2009;50:S381.
 22. Ferenci P, Scherzer TM, Kerschner H, et al. Silibinin is a potent antiviral agent in patients with chronic hepatitis C not responding to pegylated interferon/ribavirin therapy. *Gastroenterology* 2008;135:1561–1567.
 23. Sarrazin C, Kieffer TL, Bartels D, et al. Dynamic hepatitis C virus genotypic and phenotypic changes in patients treated with the protease inhibitor telaprevir. *Gastroenterology* 2007;132:1767–1777.
 24. Kieffer TL, Sarrazin C, Miller JS, et al. Telaprevir and pegylated interferon-alpha-2a inhibit wild-type and resistant genotype 1 hepatitis C virus replication in patients. *Hepatology* 2007;46:631–639.
 25. McHutchison JG, Everson GT, Gordon SC, et al. Telaprevir with peginterferon and ribavirin for chronic HCV genotype 1 infection. *N Engl J Med* 2009;360:1827–1838.
 26. Susser S, Welsch C, Wang Y, et al. Characterization of resistance to the protease inhibitor boceprevir in hepatitis C virus-infected patients. *Hepatology* 2009;50:1709–1718.
 27. Sarrazin C, Hong J, Lim S, et al. Incidence of virologic escape observed during ITMN-191 (R7227) monotherapy is genotype dependent, associated with a specific NS3 substitution, and suppressed upon combination with peginterferon alfa-2a/ribavirin. *J Hepatol* 2009;50(Suppl 1):S350.
 28. Kuloj G, Benhamou Y, Manns MP, et al. BI 201335, a potent HCV NS3 protease inhibitor in treatment-naïve and -experienced chronic HCV genotype-1 infection: genotypic and phenotypic analysis of the NS3 protease domain. *J Hepatol* 2009;50(Suppl 1):S347.
 29. Afdhal N, Godofsky E, Dienstag J, et al. Final phase I/II trial results for NM283, a new polymerase inhibitor for hepatitis C: antiviral efficacy and tolerance in patients with HCV-1 infection, including previous interferon failures. *Hepatology* 2004;40(Suppl 1):726A.
 30. Roberts SK, Cooksley G, Dore GJ, et al. Robust antiviral activity of R1626, a novel nucleoside analog: a randomized, placebo-controlled study in patients with chronic hepatitis C. *Hepatology* 2008;48:398–406.
 31. Lalezari J, Gane E, Rodriguez-Torres M, et al. Potent antiviral activity of the HCV nucleoside polymerase inhibitor R7128 with PEG-IFN and ribavirin: interim results of R7128 500mg BID for 28 days. *J Hepatol* 2008;48(Suppl 2):29A.
 32. Larrey D, Benhamou Y, Lohse AW, et al. Safety, pharmacokinetics and antiviral effect of BI 207127, a novel HCV RNA polymerase inhibitor, after 5 days oral treatment in patients with chronic hepatitis C. *J Hepatol* 2009;50(Suppl 1):S383–S384.
 33. Erhardt A, Deterding K, Benhamou Y, et al. Safety, pharmacokinetics and antiviral effect of BILB 1941, a novel hepatitis C virus RNA polymerase inhibitor, after 5 days oral treatment. *Antivir Ther* 2009;14:23–32.
 34. Troke P, Lewis M, Simpson P, et al. Genotypic characterisation of HCV NS5B following 8-day monotherapy with the polymerase inhibitor PF-00868554 in HCV-infected subjects. *J Hepatol* 2009;50:S351.
 35. Jacobson I, Pockros P, Lalezari J, et al. Antiviral activity of filibuvir in combination with pegylated interferon alfa-2a and ribavirin for 28 days in treatment-naïve patients chronically infected with HCV genotype 1. *J Hepatol* 2009;50(Suppl 1):S382–S383.
 36. Lawitz E, Rodriguez-Torres M, DeMicco M, et al. Antiviral activity of ANA598, a potent non-nucleoside polymerase inhibitor, in chronic hepatitis C patients. *J Hepatol* 2009;50(Suppl 1):S384.
 37. Villano S, Howe A, Raible D, et al. Analysis of HCV NS5B genetic variants following monotherapy with HCV-796, a non-nucleoside polymerase inhibitor, in treatment-naïve HCV-infected patients. *Hepatology* 2006;44(Suppl 1):607A–608A.
 38. Kneteman NM, Howe AY, Gao T, et al. HCV796: a selective nonstructural protein 5B polymerase inhibitor with potent anti-hepatitis C virus activity in vitro, in mice with chimeric human livers, and in humans infected with hepatitis C virus. *Hepatology* 2009;49:745–752.
 39. McCown MF, Rajyaguru S, Kular S, et al. GT-1a or GT-1b subtype-specific resistance profiles for hepatitis C virus inhibitors telaprevir and HCV-796. *Antimicrob Agents Chemother* 2009;53:2129–2132.
 40. Bartels DJ, Zhou Y, Zhang EZ, et al. Natural prevalence of hepatitis C virus variants with decreased sensitivity to NS3.4A protease inhibitors in treatment-naïve subjects. *J Infect Dis* 2008;198:800–807.
 41. Kuntzen T, Timm J, Berical A, et al. Naturally occurring dominant resistance mutations to hepatitis C virus protease and polymerase inhibitors in treatment-naïve patients. *Hepatology* 2008;48:1769–1778.
 42. Kim AY, Timm J, Nolan BE, et al. Temporal dynamics of a predominant protease inhibitor-resistance mutation in a treatment-naïve, hepatitis C virus-infected individual. *J Infect Dis* 2009;199:737–741.
 43. Colson P, Brouk N, Lembo F, et al. Natural presence of substitution R155K within hepatitis C virus NS3 protease from a treatment-naïve chronically infected patient. *Hepatology* 2008;47:766–767.
 44. Rodriguez-Frias F, Cubero M, Esteban JI, et al. Detection of clinically relevant minor HCV and HBV antiviral-resistant mutants in treatment-naïve patients using ultra-deep pyrosequencing. *J Hepatol* 2009;50:S130.
 45. Le Pogam S, Seshadri A, Kosaka A, et al. Existence of hepatitis C virus NS5B variants naturally resistant to non-nucleoside, but not to nucleoside, polymerase inhibitors among untreated patients. *J Antimicrob Chemother* 2008;61:1205–1216.

46. Marquis M, Deterding K, Erhardt A, et al. Genotypic and phenotypic analysis of hepatitis C virus NS5B polymerase variants to BILB1941 inhibition. *Hepatology* 2008;48(Suppl):1159A.
47. McKeage K, Perry CM, Keam SJ. Darunavir: a review of its use in the management of HIV infection in adults. *Drugs* 2009;69:477–503.
48. Ghany MG, Doo EC. Antiviral resistance and hepatitis B therapy. *Hepatology* 2009;49:S174–S184.
49. Ozeki I. Safety and antiviral activity of monotherapy with MP-424 for 24 weeks in naive patients with chronic hepatitis c virus genotype 1b infection. *J Hepatol* 2009;50:S350.
50. Hopkins S, Heuman D, Gavis E, et al. Safety, plasma pharmacokinetics, and anti-viral activity of SCY-635 in adult patients with chronic hepatitis C virus infection. *J Hepatol* 2009;50:S36.
51. Coelmont L, Gallay P, Bobardt M, et al. Particular in vitro anti-HCV activities and resistance profile of the cyclophilin inhibitor debio 025. *J Hepatol* 2009;50:S36.
52. Sarrazin C, Rouzier R, Wagner F, et al. SCH 503034, a novel hepatitis C virus protease inhibitor, plus pegylated interferon alpha-2b for genotype 1 nonresponders. *Gastroenterology* 2007;132:1270–1278.
53. Marcellin P, Forns X, Goeser T, et al. Virological analysis of patients receiving telaprevir administered q8h or q12h with peginterferon-alfa-2a or -alfa2b and ribavirin in treatment-naive patients with genotype 1 hepatitis C: study C208. *Hepatology* 2009;50(Suppl):395A.
54. Mocroft A, Phillips AN, Soriano V, et al. Reasons for stopping antiretrovirals used in an initial highly active antiretroviral regimen: increased incidence of stopping due to toxicity or patient/physician choice in patients with hepatitis C coinfection. *AIDS Res Hum Retroviruses* 2005;21:743–752.
55. Lin C, Gates CA, Rao BG, et al. In vitro studies of cross-resistance mutations against two hepatitis C virus serine protease inhibitors, VX-950 and BILN 2061. *J Biol Chem* 2005;280:36784–36791.
56. Lin K, Kwong AD, Lin C. Combination of a hepatitis C virus NS3-NS4A protease inhibitor and alpha interferon synergistically inhibits viral RNA replication and facilitates viral RNA clearance in replicon cells. *Antimicrob Agents Chemother* 2004;48:4784–4792.
57. Koev G, Dekhtyar T, Han L, et al. Antiviral interactions of an HCV polymerase inhibitor with an HCV protease inhibitor or interferon in vitro. *Antiviral Res* 2007;73:78–83.
58. Forestier N, Reesink HW, Weegink CJ, et al. Antiviral activity of telaprevir (VX-950) and peginterferon alfa-2a in patients with hepatitis C. *Hepatology* 2007;46:640–648.
59. Hezode C, Forestier N, Dusheiko G, et al. Telaprevir and peginterferon with or without ribavirin for chronic HCV infection. *N Engl J Med* 2009;360:1839–1850.
60. Schiff E, Poordard F, Jacobson I, et al. Boceprevir combination therapy in null responders: response dependent on interferon responsiveness. *J Hepatol* 2008;48(Suppl 2):46A.
61. Kwo P, Lawitz E, McCone J, et al. HCV SPRINT-1 final results: SVR 24 from a phase 2 study of boceprevir plus peginterferon alfa-2b/ribavirin in treatment-naive subjects with genotype-1 chronic hepatitis C. *J Hepatol* 2009;50(Suppl 1):S4.
62. Pockros PJ, Nelson D, Godofsky E, et al. R1626 plus peginterferon alfa-2a provides potent suppression of hepatitis C virus RNA and significant antiviral synergy in combination with ribavirin. *Hepatology* 2008;48:385–397.
63. Olsen DB, Carroll SS, Handt L, et al. HCV antiviral activity and resistance analysis in chronically infected chimpanzees treated with NS3/4A protease and NS513 polymerase inhibitors. *J Hepatol* 2007;46:S298.
64. Gane EJ, Roberts SK, Stedman C, et al. First-in-man demonstration of potent antiviral activity with a nucleoside polymerase (R7128) and protease (R7227/ITMN-191) inhibitor combination in HCV: safety, pharmacokinetics, and virologic results from INFORM-1. *J Hepatol* 2009;50(Suppl 1):S380.
65. Curry S, Qiu P, Tong X. Analysis of HCV resistance mutations during combination therapy with protease inhibitor boceprevir and PEG-IFN alpha-2b using TaqMan mismatch amplification mutation assay. *J Virol Methods* 2008;153:156–162.
66. Lin C, Lin K, Luong YP, et al. In vitro resistance studies of hepatitis C virus serine protease inhibitors, VX-950 and BILN 2061: structural analysis indicates different resistance mechanisms. *J Biol Chem* 2004;279:17508–17514.
67. Tong X, Bogen S, Chase R, et al. Characterization of resistance mutations against HCV ketoamide protease inhibitors. *Antiviral Res* 2008;77:177–185.
68. Qiu P, Sanfiorenzo V, Curry S, et al. Identification of HCV protease inhibitor resistance mutations by selection pressure-based method. *Nucleic Acids Res* 2009;37:e74.
69. Qi X, Bae A, Liu S, et al. Development of a replicon-based phenotypic assay for assessing the drug susceptibilities of HCV NS3 protease genes from clinical isolates. *Antiviral Res* 2009;81:166–173.
70. Ludmerer SW, Graham DJ, Patel M, et al. A transient cell-based phenotypic assay for hepatitis C NS3/4A protease: application to potency determinations of a novel macrocyclic inhibitor against diverse protease sequences isolated from plasma infected with HCV. *J Virol Methods* 2008;151:301–307.
71. Lamarre D, Anderson PC, Bailey M, et al. An NS3 protease inhibitor with antiviral effects in humans infected with hepatitis C virus. *Nature* 2003;426:186–189.
72. Hinrichsen H, Benhamou Y, Wedemeyer H, et al. Short-term antiviral efficacy of BILN 2061, a hepatitis C virus serine protease inhibitor, in hepatitis C genotype 1 patients. *Gastroenterology* 2004;127:1347–1355.
73. Reiser M, Hinrichsen H, Benhamou JP, et al. Antiviral efficacy of NS3-serine protease inhibitor BILN-2061 in patients with chronic genotype 2 and 3 hepatitis C. *Hepatology* 2005;41:832–835.
74. Zhou Y, Bartels DJ, Hanzelka BL, et al. Phenotypic characterization of resistant Val36 variants of hepatitis C virus NS3-4A serine protease. *Antimicrob Agents Chemother* 2008;52:110–120.
75. Zhou Y, Muh U, Hanzelka BL, et al. Phenotypic and structural analyses of hepatitis C virus NS3 protease Arg155 variants: sensitivity to telaprevir (VX-950) and interferon alpha. *J Biol Chem* 2007;282:22619–22628.
76. Welsch C, Domingues FS, Susser S, et al. Molecular basis of telaprevir resistance due to V36 and T54 mutations in the NS3-4A protease of the hepatitis C virus. *Genome Biol* 2008;9:R16.
77. Forestier N, Susser S, Welker MW, et al. Long term follow-up of patients previously treated with telaprevir. *Hepatology* 2008;48(Suppl):760A.
78. Shiffman ML, Berg T, Poordard F, et al. A study of telaprevir combined with peginterferon-alfa-2a and ribavirin in subjects with well-documented non-response or relapse after previous peginterferon-alfa-2a and ribavirin treatment: interim analysis. *Hepatology* 2008;48(Suppl):1135A–1136A.
79. Zeuzem S, Sarrazin C, Rouzier R, et al. Anti-viral activity of SCH 503034, a HCV protease inhibitor, administered as monotherapy in hepatitis C genotype 1 (HCV-1) patients refractory to pegylated interferon (PEG-IFN-alfa). *Hepatology* 2005;42(Suppl 1):233A.
80. Susser S, Forestier N, Welker MW, et al. Detection of resistant variants in the hepatitis C virus NS3 protease gene by clonal sequencing at long-term follow-up in patients treated with boceprevir. *J Hepatol* 2009;50:S7.
81. Forestier N, Larrey D, Guyader D, et al. Treatment of chronic hepatitis C virus (HCV) genotype 1 patients with the NS3/4A

- protease inhibitor ITMN-191 leads to rapid reductions in plasma HCV RNA: results of a phase 1b multiple ascending dose study. *Hepatology* 2008;48(Suppl):1132A.
82. Manns MP, Reesink HW, Moreno C, et al. Safety and antiviral activity of TMC435350 in treatment-naïve genotype 1 HCV infected patients. *Hepatology* 2008;48(Suppl):1023A.
 83. Manns MP, Bourliere M, Benhamou Y, et al. Safety and antiviral activity of BI201335, a new HCV NS3 protease inhibitor, in treatment-naïve patients with chronic hepatitis C genotype-1 infection given as monotherapy and in combination with peginterferon alfa 2a and ribavirin. *Hepatology* 2008;48(Suppl):1133A.
 84. Reesink H, Bergmann J, de Bruijne J, et al. Safety and antiviral activity of SCH900518 administered as monotherapy and in combination with peginterferon alfa-2b to naïve and treatment-experienced HCV-1 infected patients. *J Hepatol* 2009;50(Suppl 1):S35.
 85. Lawitz EJ, Sulkowski MS, Jacobson IM, et al. Safety, tolerability and antiviral activity of MK-7009, a novel inhibitor of the hepatitis C virus NS3/4A protease, in patients with chronic HCV genotype 1 infection. *Hepatology* 2008;48(Suppl):403A.
 86. Pasquinelli C, Eley T, Villegas C, et al. Safety, tolerability, pharmacokinetics and antiviral activity following single- and multiple-dose administration of BMS-650032, a novel HCV NS3 inhibitor, in subjects with chronic genotype 1 HCV infection. *Hepatology* 2009;50(Suppl):225.
 87. Hotho M, deBruijne J, O'Farrell A, et al. Accelerated clinical trial design to assess the safety, tolerability and anti-viral activity of PHX1766, a novel HCV NS4/4A protease inhibitor, in healthy volunteers and chronic hepatitis C patients. *Hepatology* 2009;50(Suppl):1576.
 88. Seiwert SD, Hong J, Lim SR, et al. Sequence variation of NS3/4A in HCV replicons exposed to ITMN-191 concentrations encompassing those likely to be achieved following clinical dosing. *J Hepatol* 2007;46(Suppl 1):244A.
 89. Manns M, Reesink H, Moreno C, et al. OPERA-1 trial: interim analysis of safety and antiviral activity of TMC435 in treatment-naïve genotype 1 HCV patients. *J Hepatol* 2009;50(Suppl 1):S7.
 90. Ali S, Leveque V, Le Pogam S, et al. Selected replicon variants with low level in vitro resistance to the hepatitis C virus NS5B polymerase inhibitor PSI-6130 lack cross-resistance with R1479. *Antimicrob Agents Chemother* 2008;52:4356–4369.
 91. Reddy R, Rodriguez-Torres M, Gane E, et al. Antiviral activity, pharmacokinetics, safety and tolerability of R7128, a novel nucleoside HCV RNA polymerase inhibitor, following multiple, ascending, oral doses in patients with HCV genotype 1 infection who have failed prior interferon therapy. *Hepatology* 2007;46(Suppl):862–863A.
 92. Rodriguez-Torres M, Lawitz E, Flach S, et al. Antiviral activity, pharmacokinetics, safety, and tolerability of PSI-7851, a novel nucleotide polymerase inhibitor for HCV, following single and 3 day multiple ascending oral doses in healthy volunteers and patients with chronic HCV infection. *Hepatology* 2009;50(Suppl):LB17.
 93. Lalezari J, Asmuth D, Casiro A, et al. Antiviral activity, safety and pharmacokinetics of IDX184, a liver-targeted nucleotide HCV polymerase inhibitor, in patients with chronic hepatitis C. *Hepatology* 2009;50(Suppl):LB18.
 94. Bavisotto L, Wang CC, Jacobson IM, et al. Antiviral, pharmacokinetic and safety data for GS-9190, a non-nucleoside HCV NS5B polymerase inhibitor in a phase-1 trial in HCV genotype 1 infected subjects. *Hepatology* 2007;46(Suppl 1):255A.
 95. Brainard DM, Anderson MS, Petry A, et al. Safety and antiviral activity of NS5B polymerase inhibitor MK-3281, in treatment-naïve genotype 1a, 1b and 3 HCV-infected patients. *Hepatology* 2009;50(Suppl):1568.
 96. Kukulj G, McGibbon GA, McKercher G, et al. Binding site characterization and resistance to a class of non-nucleoside inhibitors of the hepatitis C virus NS5B polymerase. *J Biol Chem* 2005;280:39260–39267.
 97. Tomei L, Altamura S, Bartholomew L, et al. Mechanism of action and antiviral activity of benzimidazole-based allosteric inhibitors of the hepatitis C virus RNA-dependent RNA polymerase. *J Virol* 2003;77:13225–13231.
 98. Hammond JL, Rosario MC, Wagner F, et al. Antiviral activity of the HCV polymerase inhibitor PF-00868554 administered as monotherapy in HCV genotype 1 infected subjects. *Hepatology* 2008;48(Suppl):1024A–1025A.
 99. Shi ST, Herlihy KJ, Graham JP, et al. Preclinical characterization of PF-00868554, a potent nonnucleoside inhibitor of the hepatitis C virus RNA-dependent RNA polymerase. *Antimicrob Agents Chemother* 2009;53:2544–2552.
 100. Cooper C, Larouche R, Bourgault B, et al. Safety, tolerability and pharmacokinetics of the HCV polymerase inhibitor VCH-222 following single dose administration in healthy volunteers and antiviral activity in HCV-infected individuals. *J Hepatol* 2009;50: S342.
 101. Lawitz E, Cooper C, Rodriguez-Torres M, et al. Safety, tolerability and antiviral activity of VCH-916, a novel non-nucleoside HCV polymerase inhibitor in patients with chronic HCV genotype 1 infection. *J Hepatol* 2009;50(Suppl 1):S37.
 102. Nicolas O, Boivin I, Berneche-D'Amours A, et al. Genotypic and phenotypic analysis of HCV NS5B variants selected from patients treated with VCH-916. *J Hepatol* 2009;50:S349.
 103. Thompson PA, Patel R, Showalter RE, et al. In vitro studies demonstrate that combinations of antiviral agents that include HCV polymerase inhibitor ANA598 have the potential to overcome viral resistance. *Hepatology* 2008;48(Suppl):1164A.
 104. Howe AY, Cheng H, Johann S, et al. Molecular mechanism of hepatitis C virus replicon variants with reduced susceptibility to a benzofuran inhibitor, HCV-796. *Antimicrob Agents Chemother* 2008;52:3327–3338.
 105. Shih I, Vliegen I, Peng B, et al. Mechanistic characterization of GS-9190, a novel non-nucleoside inhibitor of HCV NS5B polymerase with potent antiviral activity and a unique mechanism of action. *Hepatology* 2007;46(Suppl 1):859A.
 106. Rodriguez-Torres M, Lawitz E, Cohen D, et al. Treatment-naïve, HCV genotype 1-infected subjects show significantly greater HCV RNA decreases when treated with 28 days of ABT-333 plus peginterferon and ribavirin compared to peginterferon and ribavirin alone. *Hepatology* 2009;50(Suppl):5A.
 107. Koev G, Mondal R, Beyer J, et al. Characterization of resistance mutations selected in vitro by the non-nucleoside HCV polymerase inhibitors ABT-333 and ABT-072. *J Hepatol* 2009;50: S346–S347.
 108. Nettles R, Chien C, Chung E, et al. BMS-790052 is a first-in-class potent hepatitis C virus NS5A inhibitor for patients with chronic HCV infection: results from a proof-of-concept study. *Hepatology* 2008;48(Suppl):1025A.
 109. Tellinghuisen TL, Marcotrigiano J, Rice CM. Structure of the zinc-binding domain of an essential component of the hepatitis C virus replicase. *Nature* 2005;435:374–379.
 110. Chatterji U, Bobardt M, Selvarajah S, et al. The isomerase active site of cyclophilin A is critical for hepatitis C virus replication. *J Biol Chem* 2009;284:16998–17005.
 111. Mathy JE, Ma S, Compton T, et al. Combinations of cyclophilin inhibitor NIM811 with hepatitis C Virus NS3-4A Protease or NS5B polymerase inhibitors enhance antiviral activity and suppress the emergence of resistance. *Antimicrob Agents Chemother* 2008;52:3267–3275.
 112. Coelmont L, Kaptein S, Paeshuysse J, et al. Debio 025, a cyclophilin binding molecule, is highly efficient in clearing hepatitis C virus (HCV) replicon-containing cells when used alone or in combination with specifically targeted antiviral therapy for HCV (STAT-C) inhibitors. *Antimicrob Agents Chemother* 2009;53:967–976.

113. Flisiak R, Horban A, Gallay P, et al. The cyclophilin inhibitor Debio-025 shows potent anti-hepatitis C effect in patients coinfecting with hepatitis C and human immunodeficiency virus. *Hepatology* 2008;47:817–826.
114. Hopkins S, Scoreaux B, Huang Z, et al. The genetic and biochemical basis for resistance to SCY-635. *Hepatology* 2008;48(Suppl):1117A–1118A.
115. Lawitz E, Rouzier R, Nguyen T, et al. Safety and antiviral efficacy of 14 days of the cyclophilin inhibitor NIM811 in combination with pegylated interferon alpha 2a in relapsed genotype 1 HCV infected patients. *J Hepatol* 2009;50:S379.
116. Goto K, Watashi K, Inoue D, et al. Identification of cellular and viral factors related to anti-hepatitis C virus activity of cyclophilin inhibitor. *Cancer Sci* 2009;100:1943–1950.
117. Kaul A, Stauffer S, Berger C, et al. Essential role of cyclophilin A for hepatitis C virus replication and virus production and possible link to polyprotein cleavage kinetics. *PLoS Pathog* 2009;5:e1000546.
118. Korba BE, Elazar M, Lui P, et al. Potential for hepatitis C virus resistance to nitazoxanide or tizoxanide. *Antimicrob Agents Chemother* 2008;52:4069–4071.
119. Shiffman M, Ahmed A, Jacobson I, et al. Phase 2 randomized, double-blind, placebo-controlled study of nitazoxanide with peginterferon alfa-2a and ribavirin in nonresponders (NR) with chronic HCV genotype 1: week 28 interim analysis. *J Hepatol* 2009;50:S385–S386.
120. Morishima C, Shuhart MC, Wang CC, et al. Silymarin inhibits in vitro T cell proliferation and cytokine production in hepatitis C virus infection. *Gastroenterology* 2010;138:671–681.
121. Ahmed-Belkacem A, Ahnou N, Barbotte L, et al. Silibinin and related compounds are direct inhibitors of hepatitis C virus RNA-dependent RNA polymerase. *Hepatology* 2009;50(Suppl): 226.

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Conflicts of interest

The authors disclose no conflicts.

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