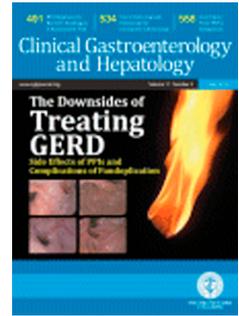


# Journal Pre-proof

Screening HIV patients at risk for NAFLD using MRI-PDFF and transient elastography: a European multicenter prospective study

Maud Lemoine, Lambert Assoumou, Pierre-Marie Girard, Marc Antoine Valantin, Christine Katlama, Stephane De Wit, Pauline Campa, Hayette Rougier, Jean-Luc Meynard, Coca Necsoi, Anja D. Huefner, Jan Van Luzen, Julian Schulze zur Wiesch, Jean-Philippe Bastard, Soraya Fellahi, Stefan Mauss, Metodi V. Stankov, Axel Baumgarten, Gerrit Post, Lawrence Serfaty, Vlad Ratziu, Yves Menu, Jerome Schlue, Pierre Bedossa, Jacqueline Capeau, Dominique Costagliola, Georg Behrens, Patrick Ingiliz, on behalf of the ANRS-ECHAM group



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# STUDY POPULATION

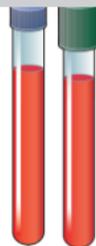
**HIV** monoinfected patients at risk of NAFLD

**≥40 years, on ART**

*MetS and/or Elevated liver enzymes and/or Lipodystrophy*



n=402, male: 85%



Standard blood tests  
Cytokines  
Biomarkers



Genetic analysis

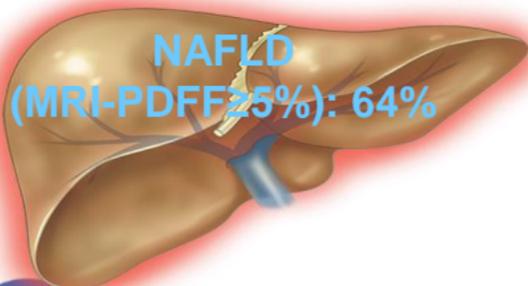


MRI-PDFF



Fibroscan-CAP

# RESULTS



**Moderate-to-severe steatosis:**

(MRI-PDFF ≥ 10%): **36%**

*Associated factors:*

*ALT, ferritin, CD4, low HDL, TG, leptin, PNPLA3 rs738409 CC*

**Advanced liver fibrosis in NAFLD:**

(≥ 9.6 kPa): **11%**

*Associated factors:*

*BMI and AST*

**CAP performance:**

**AUROC: 0.86**

**Best cut-off:**

**280 dB/m**



1 **Screening HIV patients at risk for NAFLD using MRI-PDFF and transient elastography: a European**  
2 **multicenter prospective study**

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4 Katlama<sup>4</sup>, Stephane De Wit<sup>5</sup>, Pauline Campa<sup>3</sup>, Hayette Rougier<sup>3</sup>, Jean-Luc Meynard<sup>3</sup>, Coca Necsoi<sup>5</sup>  
5 Anja D. Huefner<sup>6</sup>, Jan Van Luzen<sup>6</sup>, Julian Schulze zur Wiesch<sup>6</sup>, Jean-Philippe Bastard<sup>7,8,9,10</sup>, Soraya  
6 Fellahi<sup>7,8,10</sup>, Stefan Mauss<sup>11</sup>, Metodi V. Stankov<sup>12</sup>, Axel Baumgarten<sup>13</sup>, Gerrit Post<sup>14</sup>, Lawrence  
7 Serfaty<sup>15</sup>, Vlad Ratziu<sup>16</sup>, Yves Menu<sup>17</sup>, Jerome Schluë<sup>18</sup>, Pierre Bedossa<sup>19</sup>, Jacqueline Capeau<sup>7,8</sup>,  
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61 Abvie companies and research funding from Gilead Sciences and Abbott laboratories. DC has received  
62 speaker and consultancy fees from Janssen, Merck, MSD and Gilead Sciences companies and  
63 research funding from Merck and MSD.

64 **Abbreviations:** **AF: advanced fibrosis**, APRI: aspartate-aminotransferase to platelet ratio, AST:  
65 aspartate-aminotransferase , ART: antiretroviral treatment, AUROC: area under the receiver operating  
66 characteristic, BMI: body mass index, CAP: controlled attenuation parameter, GGT:  
67 gammaglutamyltransferase, FF: fat fraction, FLI: fatty liver index, HOMA: Homeostasis Model  
68 Assessment, NAFLD: nonalcoholic fatty liver disease, NASH: nonalcoholic steatohepatitis, LE: liver  
69 enzymes, LSM: liver stiffness measurement, MetS: metabolic syndrome, MRI-PDFF: magnetic-  
70 resonance-imaging-proton-density fat fraction, NPV: negative predictive value, PPV: positive predictive  
71 value

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74 and writing of this study.

75

76 **Abstract (254 words)**

77 **Background & aims:** Non-alcoholic fatty liver disease (NAFLD) is a growing concern in the aging HIV  
78 population. Screening for NAFLD is recommended in patients with metabolic risk factors or unexplained  
79 transaminitis. This study aimed to prospectively assess the prevalence and associated factors of liver  
80 steatosis and advanced fibrosis (AF) in HIV-monoinfected patients at risk of NAFLD.

81 **Methods:** we conducted a multicenter study in HIV-monoinfected patients, non-excessive drinkers with  
82 metabolic syndrome (MetS) and/or persistently elevated liver enzymes and/or clinical lipodystrophy. All  
83 participants had MRI-PDFF, Fibroscan@//CAP and cytokine and genetic analysis.

84 **Results:** From March 2014 to November 2015, we enrolled 442 participants and analysed 402: male  
85 (85%), median age 55 years (IQR 50-61), BMI 27.0 kg/m<sup>2</sup> (23.6-28.7), MetS (67%), CD4 cells count  
86 630/mm<sup>3</sup> (510-832). Overall 257/402 (64%) had NAFLD (MRI-PDFF≥5%). Among them, 11.3% had a  
87 liver stiffness ≥9.6 kPa, suggestive of AF. Multivariable analysis identified seven factors of steatosis:  
88 high CD4-cell count (OR:4.04 (95%CI: 1.92-8.51)), high leptin level (OR:2.12 (1.14–3.93)), non-CC  
89 PNPLA3s738409 genetic polymorphism (OR:1.92 (1.11-3.33)), low HDL (OR:1.83 (1.03-3.27)), high  
90 triglycerides (OR:1.48 (1.18-1.84)), elevated ALT (OR:1.23 (1.16-1.31)) and hyper ferritinemia (OR:1.05  
91 (1.03-1.07)). Two factors were associated with AF: high BMI (OR:1.23 (1.07-1.42), p=0.005, and  
92 elevated AST (OR:1.03 (1.01-1.05), p=0.001). Using MRI-PDFF as a reference, CAP (best cut-off 280  
93 dB/m) had good accuracy (0.86 (0.82-0.90)) for the diagnosis of moderate-to-severe steatosis.

94 **Conclusions:** In a large cohort of HIV-monoinfected patients at risk of NAFLD, steatosis is present in  
95 two third of cases and around 10% have AF. The CAP technique is accurate for screening steatosis in  
96 this population.

97

98 **Key words:** HIV, NAFLD, risk factors, steatosis, fibrosis

99

100

## 101 Introduction

102 Nonalcoholic fatty liver disease (NAFLD) encompasses a spectrum of conditions ranging from simple  
103 fatty liver (non-alcoholic fatty liver), to non-alcoholic steatohepatitis (NASH), liver fibrosis, cirrhosis and  
104 hepatocellular carcinoma.<sup>1</sup> Due to the global spread of obesity and type 2 diabetes, NAFLD is now the  
105 commonest cause of chronic liver disease<sup>2</sup> and is expected to become the first indication for liver  
106 transplantation over the next decade in developed countries.<sup>3</sup> Advanced fibrosis (AF) including cirrhosis  
107 is recognized as the most important predictor of clinical outcomes and mortality in patients with NASH<sup>4</sup>.  
108 People living with HIV (PLWH) are considered at high risk of NAFLD due to an increased prevalence of  
109 traditional metabolic risk factors and aging<sup>5,6</sup>. Life expectancy of PLWH has, indeed, greatly improved  
110 and, at present, in Western countries more than half of PLWH are more than 50 years with a high  
111 prevalence of age-related comorbidities<sup>6</sup>. Globally, the metabolic syndrome (MetS) affects 17% to 31%  
112 of PLWH<sup>11</sup> and, as a consequence, NAFLD in PLWH has now become a growing concern<sup>7</sup>. A  
113 systematic review and meta-analysis estimated the overall prevalence of NAFLD in monoinfected-  
114 PLWH as 35% (95% confidence interval (CI) 29-42) in the Western world<sup>8</sup>, but underlined the limited  
115 number of studies, often based on heterogeneous diagnostic criteria; In contrast to the non-HIV  
116 population, data on NAFLD in PLWH is scarce and heterogeneous<sup>9-16</sup> with liver histology being hardly  
117 performed in this population. This gap hampers the development of international strategies for the  
118 diagnosis and management of NAFLD in PLWH<sup>6</sup>.

119 In its 2019 guidelines, the European AIDS Clinical Society (EACS)<sup>17</sup> recommends to “*assess and*  
120 *monitor disease severity in PLWH in case of suspected NAFLD and metabolic risk factors*” and  
121 underlines the lack of data and the absence of optimal cut-off for the use of CAP, a technology widely  
122 used now in outpatient clinical services.

123 In this prospective multicenter study, using magnetic resonance imaging proton density fat fraction  
124 (MRI-PDFF)<sup>18</sup>, and liver stiffness measurement (LSM) (Fibroscan®) as the best recommended non-  
125 invasive method for the diagnosis of AF<sup>18</sup>, we aimed to assess i. the prevalence, severity and risk

126 factors of liver steatosis and AF in moninfected PLWH at risk of NAFLD and ii. the performance of the  
127 CAP technique and its best cut-off for the diagnosis of moderate-to-severe steatosis in this population.

128

## 129 **Patients and methods**

### 130 **Study population**

131 ECHAM (European Cohort on HIV, Aging and Metabolic liver disease) is a multicenter European study  
132 (7 centers: Belgium (n=1), France (n=2) and Germany (n=4)). Between March 2014 and November  
133 2015, HIV-1 infected individuals over 40 years, receiving ART for at least 5 years with HIV viral load  
134 <400 copies/mL and CD4-T cell count >100/mm<sup>3</sup>, were invited to participate to the study if they met at  
135 least one following criteria: 1- MetS defined by the 2009 international criteria<sup>19</sup> (Group 1), 2- persistently  
136 elevated liver enzymes defined by transaminases  $\geq 1.5$  upper limit of normal (ULN=35 IU/ml) and/or  
137 gammaglutamyltransferase (GGT) level  $\geq 2$ ULN (ULN=60 IU/L) on two blood samples within at least a  
138 three-month interval (Group 2), or 3- clinical lipodystrophy as previously described<sup>20</sup> (Group 3).

139 Participants were not eligible if they met one of the following criteria: positive hepatitis B or C virus  
140 serologies, coinfection with HIV-2, use of intravenous drugs within the last six months, current or past  
141 excessive alcohol intake (>30 g/day), genetic hemochromatosis, autoimmune hepatitis, primary or  
142 secondary biliary cirrhosis or cholangitis, alpha1 antitrypsin deficiency, Wilson's disease, secondary  
143 causes of NAFLD i.e ongoing prolonged steroid therapy, current therapy with amiodarone, tamoxifen,  
144 methotrexate, nifedipine, or hycanzone, history of cancer chemotherapy, short bowel syndrome,  
145 polycystic ovarian syndrome, Weber-Christian disease, active opportunistic infection except for candida  
146 oesophagitis, ongoing cancer, pregnancy, uncontrolled congestive heart failure. The study  
147 (clinicaltrial.gov NCT02093754) was approved by the national ethic committees.

### 148 **Demographic and clinical data**

149 Demographic (ethnicity), epidemiological (smoking habit, alcohol consumption, HIV transmission, year  
150 of HIV diagnosis, duration of cumulative exposure to ARV, year of ARV initiation) and clinical (BMI,  
151 waist and hip circumference, type of ARV exposure, antidiabetic, antihypertensive and hypolipidemic  
152 drugs); data were collected at enrolment in standardized electronic forms.

### 153 **Biochemical measurement**

154 Blood samples were collected after a 12-hour overnight fast for determination of liver enzymes,  
155 glucose, cholesterol, triglycerides (TG) and insulin. Measurements of circulating insulin were  
156 centralized (Architect; Abbott Laboratories, Rungis, France). Insulin resistance (IR) was assessed  
157 using the Homeostasis Model Assessment Method index (HOMA-IR) and defined as HOMA-IR  
158 index  $\geq 2.5^{21}$ .

### 159 **Serum adipokines measurement**

160 Measurement of serum adipokines was centralized in the department of biochemistry and hormonology  
161 (Tenon hospital, Paris, France): Leptin, and high sensitivity (hs) IL-6 were measured using an enzyme-  
162 linked immunosorbent assay (ELISA) (Quantikine R&D Systems, Oxford, UK). Serum adiponectin was  
163 measured by ELISA (ALPCO Salem, NH, USA) and high-sensitivity C-reactive protein (hsCRP) by  
164 immunonephelometry (IMMAGE, Beckman-Coulter).

### 165 **Genetic analysis**

166 Genomic DNA was isolated from 200  $\mu$ L of blood by QIAamp DNA Blood Mini Kit (QIAGEN, Hilden,  
167 Germany). Patatin-like phospholipase domain containing 3 (PNPLA3, rs738409 c.444C>G) and  
168 transmembrane6 superfamily 2 (TM6SF2, rs58542926 c.449C>T) polymorphisms were genotyped  
169 using TaqMan primers and probes for allelic discrimination (7500 Fast Real-Time PCR system, Applied  
170 Biosystems, Thermo Fisher Brand, Foster City, CA, USA) per the manufacturer's recommendations.

### 171 **Liver steatosis assessment**

172 For the diagnosis of NAFLD, we used MRI-PDFF, the most accurate method for detecting and  
173 quantifying hepatic steatosis according to the 2021 EASL guidelines<sup>18</sup> and which excellent performance

174 was confirmed in our population using liver histology as a reference<sup>22</sup>. We used an MRI-PDFF cut-off of  
175 5% for the definition of any degree of steatosis, and of 10% to identify moderate-to-severe liver  
176 steatosis (corresponding to  $\geq 33\%$  of hepatocytes on liver biopsy)<sup>22,23</sup>.

177 "All participants underwent a two-phase contrast hepatic MRI-PDFF with calculation of the hepatic fat  
178 fraction (FF) using a dedicated software. All machines included In Phase (IP) and Out of Phase (OP) T1  
179 imaging (T1 Gradient Echo sequences with multiple echoes and T2\* relaxometry). FF calculation in IP  
180 and OP were standardised within the study centres. For quality control, randomly selected MRI-PDFF  
181 reports were reviewed by a single highly experienced radiologist (YM) blinded to clinical patient  
182 information",

183 We also collected CAP values (dB/m) co-localized to LSM and calculated the Fatty liver index (FLI), an  
184 algorithm often used in non-HIV subjects<sup>24</sup>.

### 185 **Liver fibrosis measurement**

186 All patients had LSM, obtained in an overnight fasting state using transient elastography  
187 (Fibroscan 502, M probe, Echosens, France) and performed by ~~trained~~ experienced operators.  
188 Results were expressed in kilopascal (kPa) as the median value of 10 successful acquisitions.  
189 Failure was defined as no single successful measurement (valid shot=0) and unreliable  
190 measurement as IQR/LSM of  $>0.30$  when LSM is  $\geq 7.1\text{kPa}$ <sup>25</sup>. FIB-4 and aspartate-  
191 aminotransferase-to-platelet ratio index (APRI) were calculated as previously described<sup>18</sup>.  
192 Following the 2021 EASL recommendations on the use of non-invasive markers, we applied a cut-  
193 off 9.6kPa to detect advanced liver fibrosis<sup>18</sup>.

### 194 **Statistical analysis**

195 The patients' characteristics were described, in the entire group and according to their inclusion criteria  
196 using median and interquartile range (25<sup>th</sup>-75<sup>th</sup> percentiles) for continuous variables and number and  
197 percentage for categorical variables. We also compared characteristics of excluded and included  
198 participants. Non-parametric Kruskal-Wallis test was used to compare continuous variables between

199 the 3 groups and for categorical variables Chi-square test or Fisher's exact test when sample size was  
200 too small in any of the categories. The percentage and the associated 95%CI for steatosis and fibrosis  
201 were calculated. Univariable and multivariable logistic regression models were used to assess factors  
202 associated with moderate-to-severe steatosis defined by MRI-PDFF $\geq$ 10%, and AF defined by LSM $\geq$ 9.6  
203 kPa.<sup>18</sup> Variables with P-value  $<$ 0.20 in the univariable analysis were retained for the multivariable  
204 analyses. Because of very high number of variables and to select the most pertinent associated  
205 variables, we used a multiple imputation and bootstrap approach : we created 5 datasets in which  
206 missing data were imputed using multiple imputation approach. On each dataset, we generated 100  
207 bootstrap samples. On each of the 500 samples, a backward selection technique ( $\alpha=0.05$ ) were  
208 applied and all factors that were selected in more than 75% of the models were retained.

209 Nonparametric ROC analysis was conducted to assess the performance of CAP and FLI for the  
210 diagnosis of steatosis using MRI-PDFF as a reference. The performance was considered good for  
211 AUROC between 0.80-0.90 and excellent if  $>$ 0.90. The performances were also assessed in terms of  
212 sensitivity, specificity, positive and negative predictive values, and the likelihood ratio for a positive test  
213 result (LR+) and the likelihood ratio for a negative test result (LR-). Optimal cut-off values for CAP and  
214 FLI were selected to maximize the sum of sensitivity and specificity.

215

## 216 **Results**

### 217 **Study population**

218 461 HIV mono-infected individuals were screened, 442 met the inclusion criteria and 402 with complete  
219 metabolic and liver assessment were further analyzed (Figure 1). No statistical differences were found  
220 between excluded and analyzed participants except for HBP (43% vs 65%,  $p=0.006$ ) and ALT level  
221 (42.5 (32-65) versus 34 (24-50) IU/L,  $p=0.018$ ). Analysed participants were mainly male ( $n=340$ , 85%),  
222 median age 55 years (IQR: 50-61) and median BMI 27.0 kg/m<sup>2</sup> (23.6-28.7). All participants had  
223 controlled HIV-1 infection with a plasmatic HIV-1 RNA  $<$ 50 copies/mL -except for 11 (3%) patients, a

224 median CD4-T cell count of  $630/\text{mm}^3$  (510-832) and a median duration of cART exposure of 16 years  
225 (11-19). Most of the participants were enrolled because of MetS (group1,  $n=269$ , 67%); a minority,  
226 (group 2, 36 (8.9%)) because of persistently elevated liver enzymes without MetS and 97 (24.1%)  
227 because of clinical lipodystrophy (group 3): (74 (76.3%) patients had both lipohypertrophy and  
228 lipoatrophy, 13 (13.4%) had isolated lipoatrophy and 10 (10.4%) isolated lipohypertrophy).

229 Table 1 summarizes the characteristics of the study population and the three subgroups according to  
230 the underlying medical condition that qualified them for inclusion into the study. The participants'  
231 characteristics significantly differed between the three groups. As expected, patients with MetS had  
232 higher anthropometric parameters and metabolic disorders as compared to the two other groups.

233 All participants except 2 were assessed for PNPLA3 and TM66SF2 gene polymorphisms. PNPLA3  
234 rs738409 C/C (230 (57.5%)) and TM66SF2 rs58542926 C/C (341 (85.3%)) were the most frequent  
235 polymorphisms observed. No difference in gene polymorphism frequency was observed between the 3  
236 subgroups of patients.

### 237 **Prevalence and risk factors of liver steatosis**

238 Among the 402 study patients, 257 had a MRI-PDFF  $\geq 5\%$  and 145 MRI-PDFF  $\geq 10\%$  giving a  
239 prevalence of steatosis at any degree of 64% (95%CI: 59-69) and of moderate-to-severe steatosis of  
240 36% (95%CI: 31-41). The proportion of steatosis (MRI PDFF $\geq 10\%$ ) was the highest in patients with  
241 MetS (43%) and abnormal liver enzymes (44%) and the lowest in patients with isolated lipodystrophy  
242 (14%),  $p < 0.0001$  (Table 1).

243 As compared to patients with MRI-PDFF  $< 10\%$ , patients with MRI-PDFF  $\geq 10\%$  had a higher proportion  
244 of metabolic disorders (MetS: 79.3% *versus* 59.9%,  $p < 0.001$ ), higher CD4-T cell ( $720/\text{mm}^3$  (555-903) *vs*  
245  $597$  (488-772)  $/\text{mm}^3$ ,  $p < 0.0001$ ), higher liver transaminases (ALT (49 IU/L (34-96) *vs* 29 IU/L  $n=7.80$ ) *vs*  
246 5.10 (4.30-6.30),  $p < 0.001$ ) and a higher proportion of AF (16/109 (14.7%) *vs* 9/194 (4.6%)  $p = 0.002$ ).

247 Patients with MRI-PDFF  $\geq 10\%$  carried more frequently a single nucleotide polymorphism of PNPLA3  
248 genes compared to patients with MRI-PDFF  $< 10\%$  (75/144 *vs* 95/256,  $p = 0.0005$ ) (Suppl. Table 1).

249 In multivariable analysis, seven factors were independently associated with moderate-to-severe  
250 steatosis: CD4 cell count (OR: 4.04 (1.92-8.51)), leptin level ( $\geq 3.2$   $\mu\text{g/L}$ , OR: 2.12 (1.14–3.93)), non-  
251 CC PNPLA3s738409 genetic polymorphism (OR: 1.92 (1.11-3.33)), low HDL ( $<1$  mmol/L for men and  
252  $<1.3$  mmol/L for women) (OR: 1.83 (1.03-3.27)), triglyceride level (OR: 1.48 (1.18-1.84)), ALT level  
253 (OR: 1.23 (1.16-1.31)), ferritin level (OR: 1.05 (1.03-1.07)) (Table 2).

254

### 255 **Prevalence and risk factors of advanced liver fibrosis (AF) in patients with NAFLD**

256 Among the 257 patients with NAFLD, 63 (24.5%) had invalid LSM and 194 had valid results; as  
257 expected patients with invalid LSM had higher BMI and higher metabolic disorders compared to  
258 patients with valid LSM (data not shown).

259 Amongst 194 patients with valid LSM, 22 (11.3%) had a LSM  $\geq 9.6$  kPa suggesting AF including 11  
260 (5.7%) patients with LSM  $\geq 12.5$  kPa suggestive of cirrhosis. As compared to patients with LSM  $< 9.6$   
261 kPa, patients with AF had more metabolic disorders and higher degree of liver steatosis with higher  
262 leptin levels and leptin-to-adiponectin ratio but genetic profile (PNPLA3 and TM6SF2) was similar  
263 (Suppl. Table 2).

264 Multivariable analysis identified two factors associated with AF: high BMI (OR: 1.23 (1.07-1.42),  
265  $p=0.005$ ) and high AST level (OR: 1.02 (1.01-1.05),  $p=0.001$ ) (Table 3).

### 266 **Performance of CAP and fatty liver index (FLI) using MRI-PDFF as a reference**

267 Valid CAP values were obtained in 356 patients (89%). Using MRI-PDFF as a reference, CAP had  
268 good performance for the diagnosis of moderate-to-severe liver steatosis (MRI-PDFF  $\geq 10\%$ ) with an  
269 AUROC at 0.862 (0.823-0.901) and a best cut-off of 280 dB/m with a sensitivity of 75%, a specificity of  
270 83% and 80% of patients correctly classified. In contrast to CAP, FLI had a poor performance with an  
271 AUROC at 0.692 (0.636-0.749) (Table 4, Figure 2).

### 272 **Discussion**

273 In this multicenter European study, we conducted a comprehensive hepatic and metabolic assessment  
274 of HIV-monoinfected patients at risk of NAFLD (i.e aged over 40 years, exposed to cART for more than

275 5 years with a MetS or persistently elevated liver enzymes or clinical lipodystrophy). We selected this  
276 population, as now recommended by the EACS guidelines<sup>17</sup>, in order to describe the burden of NAFLD  
277 in the potentially most affected PLWH who are most likely to be referred for liver assessment in clinical  
278 practice.

279 We found a high proportion of NAFLD (64%) including 36% presenting moderate-to-severe steatosis  
280 and, using liver stiffness measurement, 11% had advanced fibrosis including 6% with cirrhosis.

281 Not surprisingly, patients with MetS had the highest prevalence of NAFLD (71%) whilst patients with  
282 isolated lipodystrophy had a lower prevalence (43%) and a markedly better metabolic profile and insulin  
283 sensitivity suggesting that the dysmetabolic profile with insulin resistance observed in subjects with  
284 MetS, plays a key role in the development of NAFLD, as observed in the general population<sup>1</sup>.

285 In addition to the classic metabolic parameters associated with liver steatosis (triglycerides, low HDL,  
286 ferritin and leptin levels), we identified ALT level and CD4-T cell count as associated factors of liver  
287 steatosis. The role of an improved immune system in this population as a risk factor of liver steatosis  
288 has been previously suggested<sup>8,14</sup>. This finding could reflect an immunological phenomenon or could  
289 mark a “healthier” general condition or the contribution of modern HIV drugs (e.g., integrase inhibitors)  
290 but remains to be further evaluated. We did not find any association between ART class and CD4-T cell  
291 levels.

292 We did not find association between a specific HIV drug class and liver steatosis in our study and we  
293 were unable to collect the duration of cumulative drug exposure to each specific ARV agent. Indeed,  
294 our study enrolled PLWH treated for more than 10 years with different combinations and dosages.  
295 However, discrepant results have been previously reported.<sup>14</sup> In a Danish study, exposure to an  
296 integrase inhibitor has been also identified as a risk factor of moderate-to-severe liver steatosis  
297 measured by CT-scan in consecutive PLWH.<sup>15</sup> Other studies, however, did not report association  
298 between exposure to NRTI or protease inhibitors (PIs) and liver steatosis.<sup>11</sup>

299 As described in non-HIV patients<sup>26</sup>, we found an association between PNLPL3A variants and the  
300 presence of moderate-to-severe liver steatosis. In monoinfected PLWH, only one study analyzed the  
301 association between PNLPLA3 variants and liver steatosis in 62 patients and observed that a single  
302 nucleotide polymorphism (rs738409;C>G) was associated with the degree of liver steatosis and ALT  
303 level<sup>16</sup>.

304 As MRI-PDFF is unlikely to be broadly used in routine HIV care, we validated the accuracy of CAP  
305 against MRI-PDFF for the detection of liver steatosis. The diagnostic performance of CAP in HIV-  
306 NAFLD patients, has been assessed in only one study conducted in 70 HIV-monoinfected subjects at  
307 risk of NAFLD<sup>27</sup>. As recently mentioned<sup>6,17</sup> the best cut-off of CAP needs to be confirmed. In our study,  
308 we found a very good performance of CAP for the diagnosis of moderate-to-severe steatosis and a best  
309 cut-off of 280 dB/m, close to the 285 cut-off proposed by Ajmera et al. and the 275 cut-off recently  
310 proposed by the EASL in non-HIV subjects with NAFLD<sup>18</sup>.

311 The high rate of liver steatosis found in our study may be alarming. Indeed, the results of a longitudinal  
312 study conducted in Canada in monoinfected-PLWH found that hepatic steatosis at any degree was an  
313 independent factor of liver fibrosis progression using Fibroscan®.<sup>12</sup> The long-term follow-up of our  
314 participants will be important to identify additional risk factors of liver disease progression in this  
315 population.

316 In our study, we observed advanced liver fibrosis in 11% of study subjects and BMI and AST level were  
317 identified as independent factors. This finding suggests that, as observed in non-HIV patients, obesity  
318 plays a key role in liver fibrogenesis in this population and clinical management of HIV-monoinfected  
319 patients should focus on life style changes and weight loss, as suggested in non-HIV individuals with  
320 NAFLD<sup>1</sup>. However, lean NAFLD is also frequently observed in HIV patients (24% according to a recent  
321 study<sup>28</sup>) with a risk of AF, suggesting that BMI might not be the cornerstone of NAFLD in this  
322 population.

323 Our study has strengths and limitations. To the best of our knowledge, it is the largest European  
324 multicenter study on NAFLD in monoinfected-PLWH at-risk for metabolic disorders. The long-term  
325 follow-up of these patients will fill knowledge gaps on liver disease progression in this population.<sup>6</sup> We  
326 collected a large amount of data and were able to analyze the contribution of metabolic, inflammatory  
327 and genetic markers in NAFLD in this population. We analysed mainly males and well-controlled HIV on  
328 long-term ART, whose profile is not representative of the general HIV-monoinfected population.  
329 Therefore, our rate of liver steatosis might be higher than that reported in unselected PLWH. In a large  
330 Canadian routine screening study of HIV-monoinfected patients (n=541), Pembroke et al. also reported  
331 35% of liver steatosis using the CAP technique ( $\geq 248$  dB/m) and 19% rate of liver fibrosis (LSM $\geq 7.2$   
332 kPa)<sup>12</sup>. Similar results were reported in a Brazilian study using the CAP technique (cut-off 248 dB/m)<sup>14</sup>  
333 while a recent Danish analysis using CT-scan, found a prevalence of moderate-to-severe steatosis of  
334 only 8.6%<sup>15</sup>. Finally, we were unable to assess the proportion of patients with NASH since its diagnosis  
335 requires liver biopsy, an invasive procedure, not widely accepted by the patients<sup>22</sup>.

336 In conclusion, when applying the EACS recommendations on the largest multicentre European cohort  
337 of HIV-NAFLD, our study confirms that in aging PLWH with metabolic disorders (MetS and/or  
338 lipodystrophy) and/or abnormal liver enzymes, the proportion of NAFLD (64%) and advanced fibrosis  
339 (11%) is high. One third had moderate-to-severe liver steatosis (MRI-PDFF $\geq 10\%$ ), which could be  
340 easily identified using CAP (best cut-off 280 dB/m). PLWH with high BMI and AST level should be of  
341 particular attention for liver fibrosis screening and monitoring. In the absence of validated drugs for the  
342 treatment of NAFLD in HIV-monoinfected, our data confirms that the control of metabolic disorders in  
343 this population is critical although genetic factors also play a role in the development of NALD as  
344 observed in non-HIV subjects.

## 345 **Legends**

346 **Figure 1:** study flow diagram

347 **Figure 2:** Performance of CAP and FLI for the diagnosis of moderate to severe liver steatosis using  
 348 MRI -PDFF as reference

349 **Table 1.** Characteristics of the study population

350 **Table 2:** Factors associated with moderate to severe liver steatosis (MRI-PDFF >10%)

351 **Table 3:** Factors associated with advanced liver fibrosis as defined by LSM $\geq$  9.6kPa

352 **Table 4:** Performance of CAP method and Fatty Liver Index (FLI) for the diagnosis of liver steatosis  
 353 using MRI- PDFF as a gold standard.

## 354 References

- 355 1. European Association for the Study of the L, European Association for the Study of D, European  
 356 Association for the Study of O. EASL-EASD-EASO Clinical Practice Guidelines for the management of non-  
 357 alcoholic fatty liver disease. *J Hepatol* 2016; **64**(6): 1388-402.
- 358 2. Younossi Z, Anstee QM, Marietti M, et al. Global burden of NAFLD and NASH: trends, predictions,  
 359 risk factors and prevention. *Nature Reviews Gastroenterology & Hepatology* 2017; **15**: 11.
- 360 3. Paik JM, Golabi P, Younossi Y, Mishra A, Younossi ZM. Changes in the Global Burden of Chronic  
 361 Liver Diseases From 2012 to 2017: The Growing Impact of NAFLD. *Hepatology* 2020; **72**(5): 1605-16.
- 362 4. Singh S, Allen AM, Wang Z, Prokop LJ, Murad MH, Loomba R. Fibrosis progression in nonalcoholic  
 363 fatty liver vs nonalcoholic steatohepatitis: a systematic review and meta-analysis of paired-biopsy studies. *Clin*  
 364 *Gastroenterol Hepatol* 2015; **13**(4): e1-9; quiz e39-40.
- 365 5. Verna EC. Non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in patients with HIV.  
 366 *Lancet Gastroenterol Hepatol* 2017; **2**(3): 211-23.
- 367 6. Lake JE, Overton T, Naggie S, et al. Expert Panel Review on Nonalcoholic Fatty Liver Disease in  
 368 Persons With Human Immunodeficiency Virus. *Clin Gastroenterol Hepatol* 2020.
- 369 7. Nguyen KA, Peer N, Mills EJ, Kengne AP. A Meta-Analysis of the Metabolic Syndrome Prevalence in  
 370 the Global HIV-Infected Population. *PLoS One* 2016; **11**(3): e0150970.
- 371 8. Maurice JB, Patel A, Scott AJ, Patel K, Thursz M, Lemoine M. Prevalence and risk factors of non-  
 372 alcoholic fatty liver disease in HIV-monoinfection: a systematic review and meta-analysis. *AIDS* 2017.
- 373 9. Lombardi R, Sambatakou H, Mariolis I, Cokkinos D, Papatheodoridis GV, Tsochatzis EA. Prevalence  
 374 and predictors of liver steatosis and fibrosis in unselected patients with HIV mono-infection. *Dig Liver Dis*  
 375 2016.
- 376 10. Guaraldi G, Squillace N, Stentarelli C, et al. Nonalcoholic fatty liver disease in HIV-infected patients  
 377 referred to a metabolic clinic: prevalence, characteristics, and predictors. *Clin Infect Dis* 2008; **47**(2): 250-7.
- 378 11. Vuille-Lessard E, Lebouche B, Lennox L, et al. nonalcoholic fatty liver disease diagnosed by transient  
 379 elastography with controlled attenuation parameter in unselected HIV mono-infected patients. *AIDS* 2016.
- 380 12. Pembroke T, Deschenes M, Lebouche B, et al. Hepatic steatosis progresses faster in HIV mono-  
 381 infected than HIV/HCV co-infected patients and is associated with liver fibrosis. *J Hepatol* 2017; **67**(4): 801-8.
- 382 13. Price JC, Seaberg EC, Latanich R, et al. Risk factors for fatty liver in the Multicenter AIDS Cohort  
 383 Study. *Am J Gastroenterol* 2014; **109**(5): 695-704.

- 384 14. Perazzo H, Cardoso SW, Yanavich C, et al. Predictive factors associated with liver fibrosis and  
 385 steatosis by transient elastography in patients with HIV mono-infection under long-term combined antiretroviral  
 386 therapy. *J Int AIDS Soc* 2018; **21**(11): e25201.
- 387 15. Kirkegaard-Klitbo DM, Fuchs A, Stender S, et al. Prevalence and Risk Factors of Moderate-to-Severe  
 388 Hepatic Steatosis in Human Immunodeficiency Virus Infection: The Copenhagen Co-morbidity Liver Study. *J*  
 389 *Infect Dis* 2020; **222**(8): 1353-62.
- 390 16. Morse CG, McLaughlin M, Matthews L, et al. Nonalcoholic Steatohepatitis and Hepatic Fibrosis in  
 391 HIV-1-Monoinfected Adults With Elevated Aminotransferase Levels on Antiretroviral Therapy. *Clin Infect Dis*  
 392 2015; **60**(10): 1569-78.
- 393 17. EACS. [https://www.eacsociety.org/files/2019\\_guidelines-10.0\\_final.pdf](https://www.eacsociety.org/files/2019_guidelines-10.0_final.pdf). 2019.
- 394 18. European Association for the Study of the L, List of panel m, Berzigotti A, et al. Easl Clinical Practice  
 395 Guidelines (Cpgs) On Non-Invasive Tests For Evaluation Of Liver Disease Severity And Prognosis- 2021  
 396 Update. *J Hepatol* 2021.
- 397 19. Alberti KG, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim  
 398 statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart,  
 399 Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis  
 400 Society; and International Association for the Study of Obesity. *Circulation* 2009; **120**(16): 1640-5.
- 401 20. Duvivier C, Ghosn J, Assoumou L, et al. Initial therapy with nucleoside reverse transcriptase inhibitor-  
 402 containing regimens is more effective than with regimens that spare them with no difference in short-term fat  
 403 distribution: Hippocampe-ANRS 121 Trial. *J Antimicrob Chemother* 2008; **62**(4): 797-808.
- 404 21. Wallace TM, Matthews DR. The assessment of insulin resistance in man. *Diabet Med* 2002; **19**(7):  
 405 527-34.
- 406 22. Lemoine M, Assoumou L, De Wit S, et al. Diagnostic Accuracy of Noninvasive Markers of Steatosis,  
 407 NASH, and Liver Fibrosis in HIV-Monoinfected Individuals at Risk of Nonalcoholic Fatty Liver Disease  
 408 (NAFLD): Results From the ECHAM Study. *J Acquir Immune Defic Syndr* 2019; **80**(4): e86-e94.
- 409 23. Tang A, Tan J, Sun M, et al. Nonalcoholic fatty liver disease: MR imaging of liver proton density fat  
 410 fraction to assess hepatic steatosis. *Radiology* 2013; **267**(2): 422-31.
- 411 24. Bedogni G, Bellentani S, Miglioli L, et al. The Fatty Liver Index: a simple and accurate predictor of  
 412 hepatic steatosis in the general population. *BMC Gastroenterol* 2006; **6**: 33.
- 413 25. Boursier J, Zarski JP, de Ledinghen V, et al. Determination of reliability criteria for liver stiffness  
 414 evaluation by transient elastography. *Hepatology* 2013; **57**(3): 1182-91.
- 415 26. Rotman Y, Koh C, Zmuda JM, Kleiner DE, Liang TJ, Nash CRN. The association of genetic variability  
 416 in patatin-like phospholipase domain-containing protein 3 (PNPLA3) with histological severity of nonalcoholic  
 417 fatty liver disease. *Hepatology* 2010; **52**(3): 894-903.
- 418 27. Ajmera VH, Cachay ER, Ramers CB, et al. Optimal Threshold of Controlled Attenuation Parameter for  
 419 Detection of HIV-Associated NAFLD With Magnetic Resonance Imaging as the Reference Standard. *Clin*  
 420 *Infect Dis* 2021; **72**(12): 2124-31.
- 421 28. Cervo A, Milic J, Mazzola G, et al. Prevalence, Predictors, and Severity of Lean Nonalcoholic Fatty  
 422 Liver Disease in Patients Living With HIV *Clin Infect Dis* 2020; **71**(10):694-e701.

423

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430

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432 funding and the chief investigator of the study; with the support of HR, project manager of the study,  
433 she was responsible of the study procedure and its compliance to ethic regulations.

434 PMG,PC,MAV,VK,CN,SDW,GB,JVL,JSW,AH,SM,LS,VR,AB,PI were in charge of patient recruitment.  
435 YM was in charge of the MRI-PDF calibration and analysis. PB and JS were responsible for the  
436 histopathological analysis. JPB, SF, JC were in charge of the measurement of adipokine levels and GB  
437 and MVS of the genetic analysis. LA and DC are responsible of the data management and statistical  
438 analysis. ML,PI,LA,DC and JC drafted the manuscript. All authors read it and approved it.

439 **Data availability:** the datasets generated during the ECHAM study and analysed for the current study  
440 are available from the corresponding author and the data management team (LA and DC) on  
441 reasonable request.

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	Whole population n= 402	Patients with MetS (group 1) n=269	Abnormal LE without MetS (group 2) n= 36	Patients with isolated LipoD (group 3) n= 97	P value
Age (years), median (IQR)	55 (50-61)	57 (52-63)	52 (46-57)	52 (49-56)	<0.001
Male, n (%)	340 (85)	227 (84)	34 (94)	79 (81)	0.180
Non African ethnicity, n (%)	316 (79)	211 (78)	32 (89)	73 (75)	0.322
Clinical lipodystrophy at inclusion (%)	301 (75)	191 (71)	13 (36)	97 (100)	
MetS at inclusion (%)	269 (67)	269 (100)	0 (0)	0 (0)	
Persistent abnormal LFT at inclusion (%)	98 (24)	62 (23)	36 (100)	0 (0)	
HIV parameters					
Time since HIV diagnosis, years, years	19 (14-24)	19 (13-23)	18 (10-21)	20 (16-26)	0.137
Year of HIV diagnosis (IQR)	1995 (1991-2001)	1995 (1991-2001)	1997 (1994-2005)	1995 (1989-1999)	0.025
Year of ART initiation (IQR)	1998 (1995-2002)	1998 (1995-2002)	2001 (1997-2007)	1997 (1995-1999)	0.011
Time since ART initiation,					

<b>years</b>					
<b>Duration of the cumulative exposure to ART, years (IQR)</b>	16 (11-19)	16 (12-19)	13 (7-17)	16 (13-19)	0.008
<b>HIV RNA &gt;50 cp/mL, n (%)</b>	11 (3)	8 (3)	0 (0)	3 (3)	0.572
<b>Nadir CD4, median (IQR)</b>	184 (84-266)	179 (78-259)	213 (119-277)	194 (93-280)	0.129
<b>CD4 /mm<sup>3</sup>, median, (IQR)</b>	630 (510-832)	616 (498-814)	612 (503-844)	643 (536-882)	0.801
<b>CD4/CD8 ratio, median (IQR)</b>	0.86 (0.60-1.18)	0.82 (0.55-1.09)	0.92 (0.67-1.26)	0.98 (0.70-1.29)	0.002
<b>Metabolic parameters</b>					
<b>BMI, kg/m<sup>2</sup>, median (IQR)</b>	27.0 (23.6-28.7)	26.8 (24.6-29.2)	24.7 (23.4-26.5)	23.7 (21.6-27.0)	<0.001
<b>Obesity (BMI≥30 kg/m<sup>2</sup>), n (%)</b>	71 (18)	58 (22)	3 (8)	10 (10)	<0.001
<b>Waist circumference, cm, median (IQR)</b>	98 (92-105)	101 (95-106)	94 (92-99)	91 (86-99)	<0.001
<b>Male</b>	98 (92-104)	100 (96-107)	94 (91-98)	90 (85-97)	<0.001
<b>Female</b>	101 (93-105)	101 (94-105)	103 (94-111)	99 (92-103)	0.774
<b>Elevated BP (Systolic &gt; 140 and/or diastolic &gt;90) and/or treated HBP, n (%)</b>	262 (65)	203 (75)	14 (39)	45 (46)	<0.001
<b>Impaired fasting glycemia (≥ 5.6 mmol/L) and/or anti-diabetic treatment, n (%)</b>	151 (38)	136 (51)	6 (17)	9 (9)	<0.001

<b>Triglycerides (mmol/L)</b>	1.60 (1.10-2.50)	1.82 (1.29-2.90)	1.15 (0.80-1.70)	1.30 (0.90-1.87)	<0.001
<b>Hypertriglyceridemia (triglycerides &gt;1.7 mmol/L or treatment) n (%)</b>	192 (48)	157 (58)	7 (19)	28 (29)	<0.001
<b>HDL cholesterol levels, (mmol/L), median (IQR)</b>	1.17 (0.91-1.40)	1.10 (0.90-1.30)	1.30 (1.20-1.77)	1.29 (1-1.58)	<0.001
<b>Low HDL (&lt;1 mmol/L for men and &lt;1.3 mmol/L for women), n (%)</b>	255 (63)	210 (78)	8 (22)	37 (38)	<0.001
<b>HOMA, median (IQR)</b>	2.69 (1.73-4.61)	3.21 (2.07-5.64)	2.44 (1.46-4.22)	1.94 (1.32-2.56)	<0.001
<b>HOMA <math>\geq</math> 2.5, n (%)</b>	218 (54)	175 (65)	17 (47)	26 (27)	<0.001
<b>Glucose (mmol/L), median (IQR)</b>	5.3 (4.8-6.0)	5.5 (4.9-6.3)	5.1 (4.8-5.4)	4.9 (4.5-5.4)	<0.001
<b>Leptin (microg/L), media (IQR)</b>	5.2 (2.7-10.6)	6.1 (3.3-12.3)	4.3 (1.9-7.4)	3.5 (1.7-7.4)	<0.001
<b>Male</b>	4.4 (2.4-7.9)	5.3 (3.2-9.6)	3.7 (1.9-6.2)	2.6 (1.4-4.9)	<0.001
<b>Female</b>	23.3 (14.2-39.1)	21.5 (13.4-39.1)	27.4 (22.9-31.8)	28.2 (16.5-40.4)	0.554
<b>Adiponectin (mg/L), median (IQR)</b>	2.8 (1.9-4.0)	2.7 (1.8-3.8)	3.3 (1.7-4.7)	3.0 (2.2-4.6)	0.078
<b>Leptin/adiponectin ratio, median (IQR)</b>	2.1 (1.0-4.1)	2.5 (1.2-4.3)	1.4 (0.6-2.9)	1.2 (0.6-2.9)	<0.001

hsCRP (mg/L)	1.6 (0.7-3.3)	1.7 (0.8-3.3)	1.4 (0.6-2.9)	1.2 (0.6-2.9)	<0.001
hsIL6 (pg/ml), median (IQR)	1.7 (1.1–3.0)	1.9 (1.3-3.1)	1.5 (0.9-2.3)	1.4 (1.0-2.7)	0.0001
<b>Genetic polymorphisms (n=400)</b>					
<b>PNPLA3 rs738409</b>					0.096
<b>CC</b>	230 (57%)	155 (58%)	17(47%)	58 (61%)	
<b>CG/GG</b>	143 (35%)/28 (7%)	97 (36%)/16 (6%)	17 (47%)/2 (6%)	28 (29%)/10 (10%)	
<b>TM6SF2 rs58542926</b>					0.246
<b>CC</b>	341 (85%)	233 (87%)	29 (81%)	79(82%)	
<b>CT/TT</b>	48 (12%)/11 (3%)	25 (9%)/10 (4%)	7 (19%)/0 (0%)	16 (17%)/1 (1%)	
<b>Hepatic parameters</b>					
<b>ALT (IU/L), median (IQR)</b>	34 (24-50)	35 (24-53)	62 (43-91)	28 (19-35)	<0.001
<b>AST (IU/L), median (IQR)</b>	29 (23-37)	30 (23-40)	38 (31-54)	27 (21-32)	<0.001
<b>GGT (IU/L), median (IQR)</b>	48 (29-81)	51 (31-83)	85 (54-159)	34 (24-55)	<0.001
<b>Platelet count (/mm<sup>3</sup>), median (IQR)</b>	213 (178-253)	210 (175-253)	203 (177-235)	220 (184-258)	0.289
<b>Median MRI-PDFF, % (IQR)</b>	7.0 (2.40-12.0)	7.73 (3.90-13.90)	8.05 (2.98-15.00)	3.33 (0-7.15)	<0.001
<b>FF ≥5% on MRI-PDFF</b>	257 (64)	191 (71)	24 (67)	42 (43)	<0.001
<b>FF ≥10% n (%)</b>	145 (36)	115 (43)	16 (44)	14 (14)	<0.001
<b>CAP dB/m. median (IQR) (n=356)</b>	260 (222 – 307)	267 (234-316)	261 (227-303)	228 (200-266)	<0.001

<b>FLI, median (IQR)</b>	67.2 (43.9-84.2)	74.2 (55.5-87.9)	58.5 (46.4-71.8)	42.4 (25.6-58.3)	<0.001
<b>APRI, median (IQR)</b>	0.36 (0.26-0.53)	0.38 (0.26-0.57)	0.45 (0.35-0.69)	0.30 (0.23-0.44)	<0.001
<b>Fibroscan, median (IQR) n=303</b>	5.4 (4.4-6.8)	5.7 (4.8-7.8)	5.6 (4.4-6.2)	4.9 (4.1-5.7)	<0.001
<b>LSM &lt;8 kPa, n (%) n=303 (193/32/78)</b>	251/303 (82.8%)	146/193 (75.7)	30/32 (93.8)	75/78 (96.2)	<0.001
<b>LSM ≥12.5 kPa, n (%) n=303 (193/32/78)</b>	13/303 (4.3)	13/193 (6.7)	0 (0)	0 (0)	0.021

**Table 1.** Characteristics of the ECHAM study population (n=402)

Categorical variables are expressed as raw numbers and percentages (%), continuous variables are reported as median and 25th-75th percentiles. Abbreviations: BMI: body mass index; FF: fat fraction; MetS: metabolic syndrome; LFT: liver function test, HIV: human immunodeficiency virus; ART: antiretroviral therapy; CD: cluster of differentiation; BP: blood pressure; HDL: high-density lipoprotein; LDL: low-density lipoprotein; hsCRP: high-sensitivity C-reactive protein; hsIL: high-sensitivity interleukin; PNPLA: Patatin-like phospholipase; ALT: alanine aminotransferase, AST: aspartate aminotransferase; LE: liver enzymes, LipoD: lipodystrophy, IQR: interquartile range; FLI: Fatty Liver Index; HOMA: Homeostasis Model Assessment Method index was defined as follows: fasting insulin (mU/L) x fasting plasma glucose (mmol/L)/22.5 24. Insulin resistance was defined by a HOMA index  $\geq 2.5$ .

Variable		Univariable analysis		Multivariable analysis	
		OR (CI 95%)	p-value	OR (CI 95%)	p-value
ALT level, UI/L, per 5 units		1.23 (1.17-1.30)	<0.001	1.23 (1.16-1.31)	<0.001
CD4 T cell count per log2 unit		3.22 (1.83-5.65)	<0.001	4.04 (1.92-8.51)	<0.001
Ferritin (ng/mL), per 10 units		1.06 (1.04-1.08)	<0.001	1.05 (1.03-1.07)	<0.001
Triglycerides, mmol/L, per unit		1.54 (1.30-1.84)	<0.001	1.48 (1.18-1.84)	0.001
Leptin ( $\mu$ g/L)	<3.2	1		1	
	$\geq$ 3.2	2.68 (1.66-4.31)	<0.001	2.12 (1.14-3.93)	0.017
Low HDL level (<1 mmol/L for men and <1.3 mmol/L for women)	no	1		1	
	yes	2.39 (1.56-3.66)	<0.001	1.83 (1.03-3.27)	0.041
PNPLA3 rs738409	C/C	1		1	
	Not C/C	1.84 (1.22-2.79)	0.004	1.92 (1.11-3.33)	0.020

**Table 2:** Factors associated with moderate to severe liver steatosis (MRI-PDFF  $\geq$ 10%)

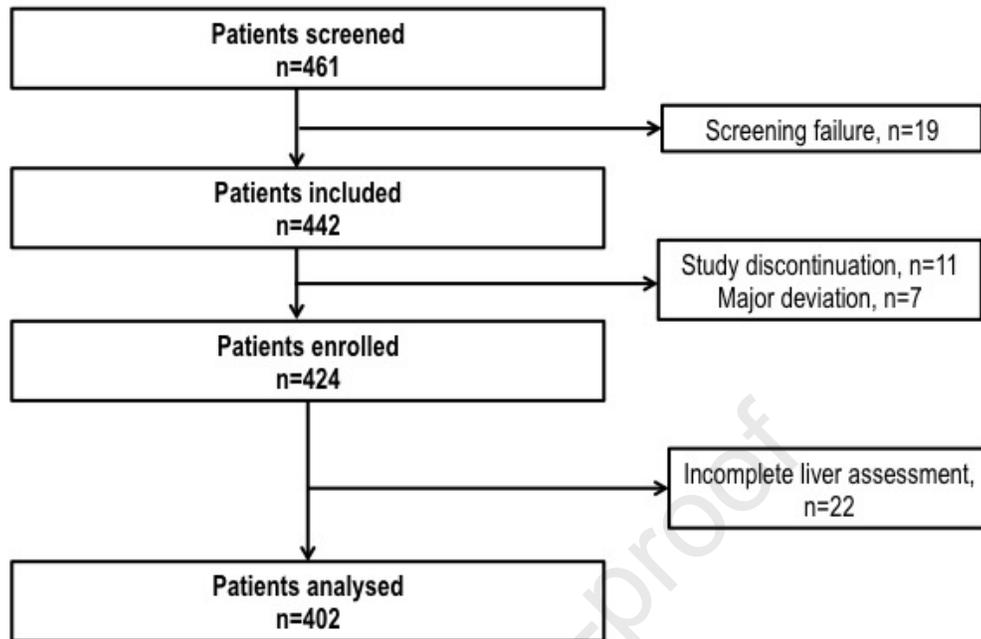
Variable	Univariable analysis	Multivariable analysis	
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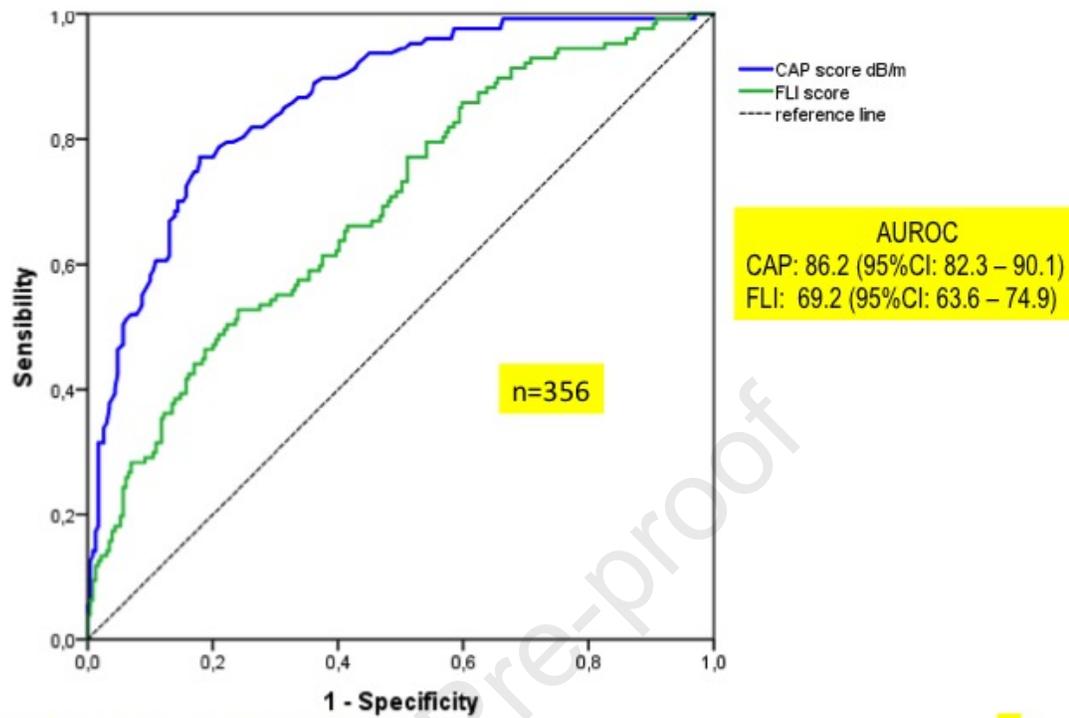
	OR (95% CI)	p-value	OR (95% CI)	P value
<b>BMI (kg/m<sup>2</sup>)</b>	1.180 (1.08-1.29)	<0.001	1.23 (1.07-1.42)	0.005
<b>AST (IU/L)</b>	1.03 (1.01-1.05)	<0.001	1.02 (1.01-1.05)	0.001

**Table 3:** Factors associated with advanced liver fibrosis as defined by LSM  $\geq$  9.6kPa in patients with MRI-PDFF  $\geq$  5%.

	Moderate to severe liver steatosis ( $\geq$ 10% MRI-PDFF)	
	CAP	FLI
<b>AUROC (95% CI)</b>	0.862 (0.823-0.901)	0.692 (0.636-0.749)
<b>Cut-off</b>	280 dB/m	75
<b>Correctly classified (%)</b>	80	68
<b>Sensitivity (%)</b>	75	57
<b>Specificity (%)</b>	83	74
<b>PPV (%)</b>	71	55
<b>NPV (%)</b>	86	75
<b>PLR</b>	4.4	2.2
<b>NLR</b>	0.3	0.6

**Table 4:** Performance of CAP method and Fatty Liver Index (FLI) for the diagnosis of liver steatosis using MRI- PDFF as a gold standard.





AUROC: area under the receiver operating characteristic  
CAP: Controlled Attenuation Parameter  
CI: confidence interval  
FLI: Fatty Liver Index  
MRI-PDFF: Magnetic resonance imaging proton density fat fraction

**What You Need to Know:**

**Background:** Aging HIV-monoinfected patients with metabolic syndrome, abnormal liver enzymes, or lipodystrophy are considered at risk of NAFLD but the severity and risk factors of liver steatosis and fibrosis remain debated in this population.

**Findings:** moderate-to-severe steatosis (MRI-PDFF $\geq$ 10%) (36%) and advanced fibrosis (Fibroscan $\geq$ 9.6kPa) (11%), are frequent. High BMI and AST are associated with advanced fibrosis.

**Implications for the patients:** Aging HIV-monoinfected patients metabolic disorders or abnormal liver enzymes should be systematically screened for liver steatosis and fibrosis.

The CAP technique (best cut-off 280 dB/m) can be used to identify patients with moderate-to-severe steatosis.